

Performance Indicators for Uranium Bioremediation in the Subsurface: Basis and Assessment

Letter Report to the Nuclear Regulatory Commission

December 15, 2006

P.E. Long S.B. Yabusaki

Hydrology Group Natural Resources Division Environmental Technology Directorate Pacific Northwest National Laboratory



DISCLAIMER

This report was prepared as an account of work sponsored by an agency of the United States Government. Neither the United States Government nor any agency thereof, nor Battelle Memorial Institute, nor any of their employees, makes any warranty, express or implied, or assumes any legal liability or responsibility for the accuracy, completeness, or usefulness of any information, apparatus, product, or process disclosed, or represents that its use would not infringe privately owned rights. Reference herein to any specific commercial product, process, or service by trade name, trademark, manufacturer, or otherwise does not necessarily constitute or imply its endorsement, recommendation, or favoring by the United States Government or any agency thereof, or Battelle Memorial Institute. The views and opinions of authors expressed herein do not necessarily state or reflect those of the United States Government or any agency thereof.

PACIFIC NORTHWEST NATIONAL LABORATORY

operated by

BATTELLE

for the

UNITED STATES DEPARTMENT OF ENERGY

under Contract DE-AC05-76RL01830

Printed in the United States of America

Available to DOE and DOE contractors from the Office of Scientific and Technical Information, P.O. Box 62, Oak Ridge, TN 37831-0062; ph: (865) 576-8401 fax: (865) 576-5728 email: reports@adonis.osti.gov

Available to the public from the National Technical Information Service,
U.S. Department of Commerce, 5285 Port Royal Rd., Springfield, VA 22161
ph: (800) 553-6847
fax: (703) 605-6900
email: orders@ntis.fedworld.gov
online ordering: http://www.ntis.gov/ordering.htm



Performance Indicators for Uranium Bioremediation in the Subsurface: Basis and Assessment

Draft Letter Report to the Nuclear Regulatory Commission

December 15, 2006

P.E. Long S.B. Yabusaki

Hydrology Group Natural Resources Division Environmental Technology Directorate Pacific Northwest National Laboratory

Table of Contents

Lis	st of Acronyms	iv
	st of Figures	
	st of Tables	
1	Introduction	1
2	Prerequisites for In Situ Bioremediation of U(VI)	
3	In Situ Bioremediation	
4	Performance Issues and Indicators	
5	Summary and Prioritization of Performance Indicators	
Re	ferences Cited	19
	bles	
	gures	

List of Acronyms

Organizational Acronyms			
BER	Office of Biological and Environmental Research in DOE/SC		
DOE	U.S. Department of Energy		
ERSD	Environmental Remediation Sciences Division of DOE/SC/BER		
ERSP	Environmental Remediation Sciences Program administered by		
DOE/SC/BER/ERSD			
NABIR Natural and Accelerated Bioremediation Research; program			
superseded by Environmental Remediation Sciences Program			
PNNL	Pacific Northwest National Laboratory		
SC	Office of Science in the DOE		
UMTRA	Uranium Mill Tailings Remedial Action program out of U.S. DOE		
	Terminology		
16S rRNA	technique to identify microorganism phylogeny by comparison to		
clone library	known gene sequences		
Ca	calcium		
DGGE	denaturing gradient gel electrophoresis; gene sequence comparison		
	technique		
DNA	deoxyribonucleic acid		
DO	dissolved oxygen		
Fe	iron		
Fe(II)	iron in the +2 oxidation state		
Fe(III)	iron in the +3 oxidation state		
ISL in situ leaching			
MCL	maximum contaminant level; drinking water standard established by		
	the U.S. Environmental Protection Agency		
Mn	manganese		
Mn(II)	manganese in the +2 oxidation state		
Mn(IV)	manganese in the +4 oxidation state		
mRNA	messenger ribonucleic acid		
ORP	oxidation reduction potential		
рН	negative log ₁₀ of the hydrogen ion activity		
PLFA	phospholipid fatty acid; analysis for microbial community structure		
	using fatty acid biomarkers		
rDNA	ribosomal DNA		
RNA ribonucleic acid			
rRNA	ribosomal RNA		
S	sulfur		
Sp.	species		
TEAP	terminal electron accepting process		
U	uranium		
U(IV)	uranium in the +4 oxidation state		
U(VI)	uranium in the +6 oxidation state		

List of Figures

- Figure 1. Dissolved uranium as a function of pH, carbonate, nitrate and sulfate ((Morrison, Spangler et al. 1995)).
- Figure 2. Water table change and associated U(VI) concentration change .
- Figure 3. Observations and modeling of DO in the subsurface.
- Figure 4. Microbial mediation of Fe(III) reduction.
- Figure 5. Borehole arrays for pilot-scale testing at the Rifle site. A. Layout of wells showing water table elevation. B. Photo of well field and sampling apparatus.
- Figure 6. Distribution of U(VI) as a function of depth during a biostimulation experiment.
- Figure 7. Passive multilevel samplers. A. Cell on support rod being lowered into monitoring well. B. MLS cells from a background well. C. MLS cells from a treatment zone well undergoing sulfate reduction.
- Figure 8. Example of heterogeneity in alluvial sediment.
- Figure 9. Plot of U(VI) concentration spatially for before and during biostimulation.

List of Tables

- Table 1. Environmentally important redox reactions in order of decreasing energy yield.
- Table 2. Prioritized information and monitoring parameters for assessment of bioremediation of U(VI) in groundwater.

Performance Indicators for Uranium Bioremediation in the Subsurface: Basis and Assessment

1 Introduction

The purpose of this letter report is to identify performance indicators for *in situ* engineered bioremediation of subsurface uranium (U) contamination. This report focuses on in situ treatment of groundwater by biostimulation of extant in situ microbial populations (see

http://128.3.7.51/NABIR/generalinfo/primers_guides/03_NABIR_primer.pdf for background information on bioremediation of metals and radionuclides). The treatment process involves amendment of the subsurface with an electron donor such as acetate, lactate, ethanol or other organic compound such that in situ microorganisms mediate the reduction of U(VI) to U(IV). U(VI) precipitates as uraninite or other insoluble U phase. Uranium is thus immobilized in place by such processes and is subject to reoxidation that may remobilize the reduced uranium. Related processes include augmenting the extant subsurface microbial populations, addition of electron acceptors, and introduction of chemically reducing materials such as zero-valent Fe. While metrics for such processes may be similar to those for in situ biostimulation, these related processes are not directly in the scope of this letter report.

The field of subsurface bioremediation has many facets and is rapidly evolving. This report, however, is limited to an overview of the underlying scientific basis for immobilization of U(VI) in groundwater via in situ biostimulation and recommendations for the monitoring of performance indicators based primarily on the authors' ongoing research in this field.

The organization of this report is based on a staged approach to full-scale field deployment that begins with a pre-existing characterization of the processes controlling uranium mobility and transport that leads to a credible conceptual model for future uranium behavior. Bench-scale studies are used to establish the proof-of-principle viability of uranium bioremediation and support the design of a field deployment strategy. A pilot-scale implementation of the uranium bioremediation strategy is used to test understanding and address uncertainties in the field conditions and behaviors that cannot adequately be addressed at the bench-scale. The elements of the approach are summarized in the following flow chart.

Characterization Prerequisites

- U distribution, form, mobility, compliance
- Hydrologic processes controlling U transport
- Geochemistry controlling U mobility



<u>Conceptual Model of Future Uranium Behavior</u> Baseline Risk Assessment for Unmitigated U Fate



Uranium Bioremediation Bench-Scale Studies

- Sequence and abundance of electron acceptors prior to onset of U reduction
- Microbial community structure / function wrt electron donor and terminal electron acceptor
- Bioavailability of terminal electron acceptors needed for U bioreduction
- Processes and kinetics controlling reoxidation and remobilization of bioreduced U
- Impact of biogeochemical reaction products on U bioreduction and subsequent reoxidation



<u>Uranium Bioremediation Conceptual Model / Strategy</u>

- Identify target microorganisms, TEAPs, electron donor
- Design electron donor concentration and delivery in context of site-specific hydrology, geochemistry, and biology
- Use simulation to assess design and long-term performance



Uranium Bioremediation Pilot-Scale Studies

- Pre-biostimulation baseline characterization
- Simulation-based monitoring/sampling scheme
- Assessment of in situ performance



Full-Scale Field Deployment

- Optimized approach
- Optimized monitoring

2 Prerequisites for In Situ Bioremediation of U(VI)

2.1 Characterization of Uranium Contamination

Uranium contamination must be sufficiently characterized in terms of source, mobility, and compliance to support selection of a remedial strategy. Furthermore, and more importantly, the attributes of this characterization must be reasonably well understood to lead to the identification of bioremediation as a cost-effective solution.

2.1.1 Uranium Inventory

The expectation is that the total uranium inventory is well-characterized in the sense that the contamination history and characterization activities have identified both current and potential future sources. Extent and concentration of uranium source are critical considerations for the bioremediation deployment strategy. Our experience has been that many groundwater uranium contamination problems have vadose zone sources. In this case, the vadose zone sources may be most active during hydrologic transients: episodic infiltration events; diurnal, seasonal, and episodic water table events. Western arid recharge rates in conjunction with uranium retardation may preclude significant contribution from recharge-driven vadose zone uranium transport. Moreover, cyclical water level fluctuations of relatively small magnitude tend to deplete a stable subsurface uranium source in the saturated zone over time frames of decades or more. However, episodic events of extended duration and/or extreme magnitude can liberate previously unleached contaminated sediments. For this reason, it might be necessary to consider augmenting bioremediation with technologies (e.g., surface barriers) that limit longer-term, recharge-driven vadose zone uranium transport.

2.1.2 Uranium Form, Concentration, and Mobility

Uranium has a broad range of mobility that is dependent on the ambient water chemistry and the surface reactivity of the sediments. We assume here that the hexavalent (+6) oxidation state [U(VI)]is the nominal valence for the bulk of the aqueous uranium species. While the hexavalent form of aqueous uranium is considered to have the most potential for transport, sorption processes for U(VI) are particularly sensitive to pH, carbonate complexation, and aqueous uranium concentration (Figure 1).

Typically, the characterization of uranium geochemistry is done in laboratory studies with contaminated and uncontaminated site sediments. Extraction procedures attempt to classify the uranium pools on sediments in terms of leachability with different extractants (e.g., labile versus nonlabile using carbonate and acid extraction). A critical issue is to differentiate the labile uranium from the rest of the sediment-associated uranium. Nonlabile uranium may be in the form of 1) sparingly soluble uranium and uranium-incorporated minerals, or 2) uranium that is not easily desorbed under ambient or engineered chemical conditions. Under circum-neutral pH and calcite-controlled carbonate chemistry in western arid sediments, we have seen the (labile) uranium partitioning coefficient, Kd (ml/g), for the less than 2 mm sediment size fraction range from less than 1 (DOE 1999) to greater than 20 (Qafoku, Zachara et al. 2005).

Another aspect of geochemical control on mobility is the reactive sediment surface environment. Although there is a body of work focused on Fe(III) minerals as the principal surface complexation site cf. (Waite, Davis et al. 1994; Payne, Davis et al. 1996), other researchers cf. (Arai, McBeath et al. 2006) have identified poorly crystalline aluminosilicate mineral as potential primary sorption surfaces. Sorption site density (e.g., surface complexation sites) is an input parameter to mechanistic sorption models that is often related to mineral surface area (Davis and Kent 1990).

The significance of the uranium concentration is also important from a compliance perspective. The drinking water MCL of 30 μ g/L may not be the regulatory compliance criteria for sites that are not drinking water sources. For example, the Old Rifle, Colorado UMTRA site cleanup criteria is 44 μ g/L. In any case, negotiated compliance standards relative to the existing contamination levels should play a role in the level of cleanup required and thus, the remediation technology selection.

2.2 Characterization of Relevant Transport Processes and Properties

In addition to the characterization of the uranium source term and geochemistry, an equally important prerequisite to the selection and design of a remediation technology is the characterization of the relevant transport processes and properties that are operative at the site for uranium. In this case, the objective is to develop an understanding of the environmental properties and process dynamics on the site that will control/influence uranium fate.

2.2.1 Hydrologic System Dynamics

As pointed out above, a focus on the groundwater system alone may ignore a significant and persistent long-term source from the vadose zone. Even if the vadose zone does not need to be explicitly addressed, seasonal water table fluctuations or influence from surface water systems (e.g., rivers, reservoirs, lakes, impoundments) should be considered in the analysis. A critical issue is the dynamics of the principal transport pathway to compliance points/surfaces, especially in the case of directional changes in the regional and local groundwater flow field.

2.2.1.1 Boundary conditions

The driving forces for the hydrologic system must be identified and their magnitudes known to be able to identify the various transport pathways and timescales.

Recharge. Spatially and temporally variable (Gee, Ward et al. 2002) recharge may be an important hydrologic component for some aquifer systems but must definitely be addressed for the case of uranium mobilization from contaminated vadose zone sediments. This is especially important when a retarded uranium front has already reached the groundwater. If the surface sites for uranium sorption are fully loaded in the system at this point, aqueous uranium can essentially travel through the vadose zone with the infiltrating water.

Recharge is the net result from a competing set of processes (e.g., precipitation, runoff, evaporation, transpiration) that vary with material properties, climate, and site-specific conditions. An inexpensive technique for an integrated estimate of recharge is chloride mass balance, which is based on concentrations of chloride dissolved in meteoric and pore water, and the volume of meteoric water. Water fluxmeters can be used for point estimates. They work well for sands, while for silts and clays the operational recharge range is above a few hundred mm/yr (Gee, Ward et al. 2002).

Water table fluctuation. Water table fluctuations are important for transferring uranium between the aquifer and vadose zone. In the case where uranium-contaminated sediments are present in the vadose zone just above the average or nominal water table, leaching of uranium by seasonally and/or episodically rising water levels can result in a source to groundwater that can be significantly larger than recharge-driven uranium transport. Furthermore, water table fluctuation combined with groundwater uranium transport can displace uranium in the lower vadose zone to down-gradient locations. This can result in a distributed source of vadose zone uranium that is activated by seasonally and/or episodically high water tables (Figure 2).

Another potential consideration for any remedial action based on lowering and/or maintaining the oxidation-reduction potential (i.e., creating/maintaining chemically reducing conditions) is the presence of a vadose zone oxygen source. This can occur when the gas phase of the vadose zone is in direct communication with atmospheric oxygen at the ground surface. Reoxidation from oxygen diffusing through the water table can inhibit the effectiveness of engineered reducing conditions. This effect can be exacerbated by gas entrapment during water table rise that provides an enhanced pathway for oxygen to be partitioned to the aqueous phase. This can be problematic if a significant part of the uranium inventory is near the water table. Figure 3 shows dissolved oxygen stratification as the result of modeling oxygen diffusion through the water table coupled with regional groundwater flow and biologically-mediated oxygen removal.

Flooding can result in an extreme water table or recharge event that may have major impacts on uranium fate and transport including impacts to natural bioreduction. One example is the Gunnison UMTRA site where flood irrigation is practiced on pastures overlying part of the uranium plume at the site. Flood irrigation appears to either have created a downward gradient that has forced the plume downward or has created a downward flux of soil organic carbon that has accelerated natural bioreduction of uranium. Depending on the interaction of floodwater with vadose zone and soil materials and the resulting geochemical or microbiological changes, flooding events could either decrease or increase uranium concentrations in a plume. If flooding is expected to occur at a site, it is particularly important to understand in advance its likely impact and devise a strategy for assessing and mitigating any anticipated increases in uranium concentration

<u>Heads</u>. Understanding the driving forces for the groundwater flow field, especially when they are transient in time and spatially complex, is important for predictive purposes. Time series of piezometric heads from an adequate distribution of monitoring wells will usually be necessary to drive a flow model. If such a data collection network does not exist from earlier monitoring, it is relatively easy to equip existing wells with hourly reading, self-contained water level monitors. In addition, new injection or monitoring wells installed for bioremediation provide opportunities to directly test prior flow models.

2.2.1.2 Vadose zone flow and transport

If the vadose zone is important as a uranium source, characterization must address the material properties controlling flow rates and directions. Characterized parameters include porosity and constants for relationships between capillary pressure, saturation, and hydraulic conductivity (e.g., van Genuchten, Brooks-Corey). Variably saturated flow modeling with these parameters, in the context of pressure head and recharge boundary conditions, will provide the basis for analyzing and predicting the vadose zone component of uranium migration. Unsaturated permeability anisotropy created by adjacent fine and coarse grained layers are particularly important in creating lateral flow in the vadose zone, possibly causing uranium contamination to appear in the groundwater offset from its location of highest concentration in the vadose zone.

2.2.1.3 Saturated flow field

The saturated flow field will probably be the principal pathway for uranium transport to a compliance point or surface. Thus, the characterization of hydraulic conductivity and porosity in the context of accurate boundary conditions is critical to the prediction of long-term transport. Key saturated hydrogeologic parameters include hydraulic conductivity, porosity, and dispersivity. Material properties important to uranium transport include grain size distribution, porosity, selected mineral abundances (e.g. clays, Fe(III) minerals) and extractable Fe(II) abundance (also important to sustaining Fereducing conditions during bioremediation).

2.2.2 Reactive Transport

2.2.2.1 Mineral Reactions

Transported uranium is typically attenuated through association with the solid phases of the porous media. This can occur through precipitation of uranium mineral or coprecipitation of uranium during formation of other secondary minerals. For the uranium concentrations and geochemical conditions that we have encountered, ambient geochemical conditions are seldom conducive to the precipitation of uranium minerals. It is typically an exotic waste form (e.g., chemical waste stream), engineered system (e.g., polyphosphate injection and resulting autunite formation), or reducing geochemical condition that results in the precipitation of uranium minerals. However, it is possible that sequestration of uranium could occur during co-precipitation events associated with changes in the oxidation-reduction potential.

2.2.2.2 Sorption Reactions

The dominant non-biotic uranium attenuation is more likely to be controlled by sorption processes. Uranium partitions between the aqueous and solid phases as a function of the reactivity of surface minerals and water chemistry. Reactivity is typically measured in experimental studies over a range of aqueous chemical conditions with field sediments from the <2 mm size fraction. Particle size distributions and mineralogical analyses of the sediments can be useful when there is significant variability in the sorption behavior.

2.2.2.3 Water Chemistry

Spatial and temporal variability in field water chemistry should be known a priori to guide the chemical conditions for the experimental studies. Concentrations of major ions [especially those important to 1) uranium sorption: carbonate, calcium; and 2) redox chemistry: dissolved oxygen (DO), nitrate, iron, manganese, sulfate, ammonium] should be complemented by measurements of pH, alkalinity, ORP, specific conductivity, and temperature collected at the same time as samples for the major ions. Dissolved organic carbon concentrations are also needed as an indicator of background electron donor support of natural bioreduction of U(VI). These measurements provide the basis for the characterization of the field water chemistry, but must be augmented by analyses for reagents used and by-products formed during biostimulation.

2.3 Conceptual Model of Future Uranium Behavior

The prerequisite characterization of uranium contamination along with relevant transport processes and properties will form the basis of a conceptual model of future uranium behavior. The extension of this conceptual model to a systematic and quantitative coupled process prediction of unmitigated uranium fate can then be used in a baseline risk assessment. The risk assessment usually drives the decision-making for engineering intervention, regulatory compliance, and rationale for specific remedial options. A key issue is uncertainty in the characterized transport processes due to inadequate sampling and monitoring. Risk of failure from the propagation of estimation errors in the process model parameters can be reduced through well thought out monitoring schemes. Where feasible, these schemes should match the time and space scales of the sampled field processes and properties, and have sufficient flexibility to address episodic or extreme events.

Understanding uranium fate and transport in terms of the uranium extent, magnitude, form, and mobility and the environmental process dynamics is critically important to the reliability of the risk assessment. Attenuation mechanisms, such as sorption, precipitation, and dilution, need to be understood in terms of known variability in hydrologic, geochemical, and biological conditions. Thus, it is important that the monitoring of performance indicators include confirmation of the processes and conditions that form the basis of the conceptual model of future uranium fate.

In summary, pre-remediation monitoring of groundwater parameters is a critical prerequisite for a mechanistically systematic understanding of site-specific uranium behavior. Targeted data should include hydraulic head (hourly water level data), U(VI), alkalinity; dissolved oxygen (DO), nitrate, Fe(II), Mn, sulfate, ammonium, pH, ORP,

specific conductivity, and temperature. These measurements will be most useful if made using event-based sampling, (e.g., low and high water table, after major storm events) as well as on a more regular frequency. Once key behaviors and the presence or absence of constituents (e.g. sulfate) are established, the analyte list can be shortened and sampling frequency reduced to capture known trends and event responses. The monitoring list above presumes the fundamental information for hydraulic properties, boundary conditions, uranium sorption, etc. are known.

3 In Situ Bioremediation

Once the decision is made for engineering intervention to achieve compliance goals, the evaluation of remediation alternatives should be based on the site conceptual model and the attributes of the various remediation technologies. Immobilization is currently the most feasible in situ approach for uranium remediation in environmental systems. In situ approaches are typically preferred over pump and treat systems because of considerations of generated waste, worker safety, and accumulation of cost over very long time horizons. To implement immobilization approaches, biological and/or chemical conditions are manipulated to drive reactions that result in the conversion of mobile aqueous U(VI), typically as uranyl ion UO₂⁺⁺ or uranyl complexes, to immobile forms. Bioremediation typically accomplishes this through the microbially-mediated reduction of aqueous U(VI) [+6 oxidation state] in porewater to immobile U(IV) phases [+4 oxidation state]. The most common U(IV) target form is the mineral, uraninite (UO₂), although a number of other minerals can occur (Burns 1999). Bioprecipitated uraninite has been shown to precipitate as nanoparticles (Suzuki, Kelly et al. 2002) and recent column studies (Komlos et al in prep.) suggest that uraninite precipitates may be transported as originally suggested by Suzuki et al (2002). Results from field studies discussed below suggest that at field scales (temporal and spatial) bioprecipitation succeeds in immobilizing U(VI) likely by a combination of aggregation of nanoparticles and attachment to mineral surfaces.

3.1 Microbially-Mediated Processes

The reduction process occurs as microorganisms consume an electron donor (e.g., organic substrates such as ethanol, acetate, lactate), which are oxidized during metabolism to gain energy and build cellular material, while transferring electrons out of their cells to terminal electron acceptors [e.g., oxygen, nitrate, Fe(III), sulfate, bicarbonate, figure 4]. The sequence of the various terminal electron accepting processes (TEAPs) is related to the energy yield from particular microorganism, electron donor, electron acceptor combinations. For typical microbial consortia, this sequence is similar to the thermodynamic redox "ladder" in aquatic chemistry: oxygen, Mn(IV), nitrate, Fe(III), sulfate, carbonate (Table 1). Uranium, in this context, is typically present in trace quantities when compared to the dominant biogeochemical conditions maintained by the background microbial consortia, major ion chemistry and primary reactive surfaces. At these trace concentrations, uranium bioreduction is not easily differentiable from the concomitant dominant TEAP. At the Rifle site, for example, acetate-oxidizing dissimilatory iron reducing bacteria (i.e., Geobacter *sp.*) are mediating uranium

bioreduction (Anderson, Vrionis et al. 2003). Conversely, ethanol-oxidizing sulfate reducing bacteria appear to be responsible for the primary uranium bioreduction observed at the Oak Ridge FRC (Scheibe, Fang et al. 2006).

3.2 Methodology Selection

The use of any redox-based technology has a number of issues to address including 1) sequence of chemical components that must be reduced before uranium bioreduction can take place, 2) bioavailability of terminal electron acceptors, 3) reoxidation and remobilization, 4) oxidizing sources, and 5) evolving reactivity. The objective is to identify a bioremediation approach that is best suited to the attributes of the uranium inventory and the environmental system.

In addition to oxygen, there are several potential oxidizing agents present in natural systems. Depending on pH, nitrate and minerals with Mn(IV) and/or Fe(III) may be the dominant oxidizer after oxygen. At circumneutral pH, the thermodynamically favored sequence would follow the redox ladder in Table 1: oxygen, nitrate, nitrite, Mn (IV), Fe (III), sulfate, carbonate. Unless there are kinetic limitations, stronger oxidizers must be depleted prior to the availability of weaker oxidizers for reduction. Thus, it may be necessary to include the removal of oxygen and nitrate in the bioremediation methodology. While this may not be a significant issue at low nitrate, suboxic sites like the Rifle site, many sites have high nitrate concentrations (e.g., Hanford, Oak Ridge) that could drastically increase the cost and complexity of bioremediation, especially where there is a continuous influx of oxygen, nitrate, and U(VI) into the treatment zone. Vadose zone sources of these oxidized components could be important if enhanced through high recharge and/or water table fluctuation. Post-biostimulation reoxidation and remobilization of uranium, which have been observed in laboratory settings (Komlos et al in prep), may also be accelerated in the field by the presence of these oxidizers.

Remobilization of sorbed, precipitated, and co-precipitated uranium may be dependent on the dissolution kinetics of the secondary minerals that incorporate or overlie the sequestered contaminants. Furthermore, secondary minerals can contribute to the long-term stability of biogenic uraninite by slowing the reoxidation process. Abdelouas (Abdelouas, Lutze et al. 1999) found that when oxidizing background conditions returned after biostimulation ended, mackinawite (FeS) that precipitated during biotransformation of U(VI) to uraninite, provided an oxygen sink that prevented the oxidation of uraninite. Changes in the structure and function of microbial community dynamics, mineral dissolution and precipitation, and biomass production at bioremediation sites are indicative of the evolving reactivity of the subsurface system and should be monitored in the context of long-term uranium mobility. It is also possible that precipitation of stable secondary phases may serve to isolate previously precipitated sorbed or reduced U, thus removing it from direct contact with pore waters. In this context it is important to note that the goal is to maintain a sufficiently low rate of remobilization such that groundwater standards are maintained.

A key consideration is thus the maintenance of the desired TEAP. At the Rifle site, the principal uranium bioreduction has been attributed to iron-reducing *Geobacter* species.

As bioavailable Fe(III) minerals were depleted near the point of acetate injection at the Rifle site, acetate-oxidizing sulfate reducers succeeded the iron-reducers. The transition to sulfate-reducing conditions was accompanied by an increase in U(VI) concentrations in the groundwater. The inefficiency of U(VI) removal from groundwater when acetate-oxidizing sulfate reducers became dominant was identified by Anderson et al. (2003) as an important consideration in the optimization of a bioremediation strategy based on this approach. Clearly, the selection and implementation of a successful bioremediation strategy requires a reasonably good understanding of the site-specific geochemical conditions and the maintenance of specific microbial populations.

3.3 Bench-Scale Testing For Proof-Of-Principle

The sensitivity of bioremediation efficacy to field attributes (e.g., contaminant inventory, material properties, biogeochemistry, hydrology) is sufficiently high that engineered solutions must be appropriately tailored. From this perspective, a necessary performance indicator is a set of bench scale tests using site materials where possible (e.g., uncontaminated and uranium-contaminated sediments, groundwater) that address proof-of-principle as well as methodology optimization (e.g., electron donor selection and delivery). Bench-scale testing can address:

- 1. Characterization of unmitigated uranium behavior in terms of variable
 - a. geologic materials
 - b. water chemistry
 - c. geochemistry
- 2. Electron donor selection; density and composition issues for delivery
- 3. Achieving onset of principal TEAP associated with uranium bioreduction
- 4. Maintaining bioreduction efficacy

A significant range of bench scale tests have already been completed and published starting with Lovley's original bottle incubations showing microbial reduction of U(VI) (Lovley, Phillips et al. 1991) and numerous others since (see bibliography). These earlier studies make it possible to perform a minimal set of new tests with site materials to achieve the objectives noted above.

Assessment of proof-of-principle needs to account for the impact of site complexity (geohydrologic, geochemical, and microbiological) on bioremediation performance. For example, low-permeability zones (silt or clay) may not only constrain flow in a system but may also slowly release U(VI) from dead-end pores, maintaining a flux of U(VI) not anticipated from consideration of coarser grain materials (LaBolle and Fogg 2001). Appropriate design of an electron donor system can address these situations (Roden and Scheibe 2005).

In many cases, the understanding of these complexities is qualitative in nature. However, considering their potential impact is still key to effective design of the bioremediation system and helps avoid surprises during implementation. Further, newly developed geophysical monitoring techniques, especially electrical methods (Williams, Kemna et al. 2006), offer promise of inexpensive, minimally invasive approaches sensitive to

geochemical heterogeneity initially and to changes in the distribution of TEAP's during bioremediation.

3.4 Conceptual Model for Uranium Bioremediation

Based on the success of the bench-scale testing and the site baseline analysis, a conceptual model for the engineered bioremediation of a particular site can be formulated. Typically, this knowledge will be incorporated into a quantitative simulation of the dominant processes operative on the site before, during, and after biostimulation. The laboratory-based parameters used in the simulations will likely deviate from those in the field, thus these analyses are preliminary in nature. The value in numerical modeling is being able to identify general responses to variations in design parameters. This will allow the assessment of implementation issues and comparison of bioremediation options in the context of the uranium inventory, environmental transport processes, and the characterized biogeochemical reactions.

3.5 Pre-Biostimulation Baseline Characterization

A key aspect of the monitoring program should be the pre-biostimulation baseline characterization that will be necessary for comparisons with monitoring performed during and after bioremediation or pilot scale tests. It is possible that much of this baseline characterization may have already been a part of earlier characterization performed in support of the risk assessment. One particularly useful consideration is a transport experiment with multiple tracers that have different diffusion characteristics. This will provide important information regarding transport (e.g., effective porosity, preferential flow, transport time scales, multiregion behavior) that will have direct implications for the design of amendment delivery. Key baseline data required include groundwater geochemistry and sediment geochemistry (see discussion above and Table 2 for details).

Point samples from groundwater and sediment may be augmented by geophysical data collection to provide a geophysical baseline prior to conducting pilot scale testing or full-scale bioremediation. Geophysical measurements (complex resistivity, self potential, cross-well radar, and cross-well seismic) provide minimally-invasive monitoring of both initial heterogeneity of redox status and microbially-mediated changes in subsurface mineralogy and redox status. Initial complex resisitivity data from the Rifle site indicate that both changes in Fe-bearing mineral coatings on detrital grains and in sulfide mineral abundance can be tracked in 3-D, enhancing our ability to interpret groundwater and sediment geochemical changes and ultimately providing a means for real-time monitoring of uranium bioremediation. While the use of self potential to identify the spatial extent of the electrochemical response to biostimulation is relatively new, the initial results have been very encouraging with excellent consistency and signal-to-noise attributes. At the Rifle site, standard surface electrode deployments are being augmented by copper electrode rings on the outside of PVC well casings to increase spatial resolution afforded by cross-borehole analyses.

3.6 Pilot Scale testing

There are many aspects of field-scale bioremediation that cannot be adequately addressed at the bench scale including: 1) > 2 mm sediment size fraction, 2) structured,

multidimensional, multiscale physical and chemical heterogeneities, 3) microbial microenvironments, 4) density effects, and 5) seasonal and episodic hydrologic transients. The tradeoff with pilot scale testing in the field is the diminished ability to control and observe the engineered reactions. For these reasons and to reduce the risk of failure, pilot-scale field tests are likely to be cost-effective. Such tests provide a bridge to the full-scale deployment of the bioremediation technology that increase understanding of the in situ behavior and allow optimization of the final approach. Monitoring of performance indicators to confirm the field viability of the bioremediation approach is particularly important at the pilot scale, since the results can be used to select the necessary and sufficient performance indicators for full-scale deployment. Again, this is likely to reduce cost of full-scale deployment.

Figure 5 shows the layout of ERSP boreholes for the Old Rifle UMTRA Site at the end of the 2004 experiment, which provides examples of pilot-scale injection and monitoring well configurations. The larger borehole array shows one row of 3 background monitoring wells, one row of 20 injection wells, and three rows of 5 treatment-zone monitoring wells. The overall size of the plot is ~20 m on a side. The smaller borehole array consists of 1 background well, 5 injection wells, and 4 treatment zone monitoring wells. Our experience at this particular site suggests that pilot scale borehole arrays will provide optimal results if they are intermediate in size between the two shown in Figure 5. For the Rifle site, this would mean ~10 m on a side with 3 background monitoring wells, ~10 injection wells, and ~8 down-gradient monitoring wells. This approach assumes that natural gradient pilot scale experiments would be performed. Alternative well designs are possible for both natural and forced gradient experiments.

4 Performance Issues and Indicators

In this section, we describe monitoring of performance indicators that will address the potential issues related to uranium bioremediation. We divide the monitoring approach into three broad groups to assess and confirm: A) uranium removal effectiveness, B) environmental transport conceptual model, and C) microbiological conceptual model. In the following section (Section 5), we provide our best estimate of the priority of the performance indicators.

A general consideration for all three groups is to provide performance indicators with regard to principal issues with engineering uranium behavior. The first is the identification of potentially uncharacterized sources (e.g., vadose zone uranium, dissolved oxygen from water table fluctuation). At the Hanford Site 300 Area for example, the largest uranium plume is associated with the most recently used surface disposal facility that operated until 1994. Recent groundwater measurements, however, have identified a uranium hot spot near a small disposal trench that was decommissioned in 1963. The site conceptual model is also being revised to reflect a chronic source of uranium in the lower part of the vadose zone that is activated during periods of high water table.

A second performance issue revealed through groundwater monitoring during bioremediation is the presence of U(VI) in the treatment zone that is not immobilized.

While this could be the result of an uncharacterized uranium source, the interest here is for zones that are not sufficiently exposed to electron donor (and associated tracer such as bromide). In early experiments at the Rifle site, a lack of mixing in injection gallery wells resulted in the delivery of electron donor that bypassed the uppermost part of the saturated zone (Figure 6). Injectate density effects apparently played a key role, but preferential flow paths were also evident as the highest acetate and tracer concentrations were found in the *second* row of monitoring wells downgradient from the injection. This situation could be further complicated by the presence of enhanced dissolved oxygen and U(VI) near the water table. A key monitoring consideration that revealed the vertical variability and stratification during biostimulation was depth-specific sampling for both groundwater (multilevel water sampling in and around the treatment zone, Figure 7) and sediments (core samples, Figure 8).

A third performance issue is the potential for changing bioremediation effectiveness. The transition from iron to sulfate reduction described earlier at the Rifle site was shown to significantly decrease the efficacy of U(VI) removal from groundwater. However, the post-biostimulation uranium behavior showed a subsequent evolution to another biogeochemical state where enhanced uranium removal took place only in the zones that experienced significant sulfate reduction. A key concept is sampling frequency based on the time-scales of the process dynamics, which maybe diurnal, seasonal, and/or episodic. Even the seasonal sampling during the spring runoff should be viewed as event-driven because the specific timing can vary from year to year.

4.1 Group 1: Uranium Removal Effectiveness

The uranium removal effectiveness group (Group 1) provides a general perspective of the uranium distribution in the subsurface. It does not provide sufficient confirmatory information for the baseline conceptual model of uranium transport and fate addressed in Group 2, or the uranium bioremediation conceptual model addressed in Group 3.

Compliance standards are typically based on the aqueous uranium concentrations. Thus, the most direct indication of compliance status will be through monitoring of groundwater for uranium concentrations. Key components of the Group 1 groundwater monitoring indicators are to establish the initial uranium (i.e., pre-biostimulation) for both the background and contaminated aquifer zones, while capturing the transient aqueous uranium entering the treatment zone. This will provide a basis for comparing uranium concentrations monitored during and after the biostimulation, and linking observed changes with the biostimulation. In some cases (e.g., Old Rifle UMTRA site), the treatment zone may be small compared to the extent of uranium contamination such that an uncontaminated up-gradient zone is not sufficiently close to the treatment zone to reasonably compare changes. In these cases, the unmitigated contaminated zone up-gradient of the treatment zone should be monitored to identify the transient water chemistry and the background geochemical condition of the sediments. See Figure 9 for a depiction of the spatial distribution of U(VI) before and during biostiumulation.

Two key facets of U(VI) concentrations are 1) the vertical distribution in the aquifer and 2) the temporal changes associated with water table fluctuations per discussions earlier in

this document. In most cases, the amount of data that can be collected in both space and time is constrained by available funds. However, it is important to note that information on the spatial and temporal distribution of U(VI) may actually save remediation costs, if it is discovered that U(VI) concentrations are restricted to the upper part of an aquifer, for example. Furthermore, we have used passive multi-level samplers that only require a single well bore [see Vrionis et al. (2005) and Figure 7] to obtain data on the depth distribution of U(VI) as well as other dissolved constituents in groundwater.

Sampling frequency is a crucial parameter that needs to be adaptively established based on events suspected of impacting U(VI) (e.g. water table rise) or on time frames previously observed for response of U(VI) concentrations to biostimulation (7 to 10 days). We typically conduct sampling at frequencies ranging from two times per week at the outset of electron donor amendment to monthly during post-amendment monitoring. After event impacts are well established, monitoring could occur less frequently. It should also be possible to link sampling to near-continuous geophysical monitoring, possibly limiting sampling to real-time changes in geophysical response (e.g. a decrease in self-potential voltage).

Concentrations of U(VI) in groundwater should be complemented by uranium extractions performed on sediments sampled before and after primary biostimulation. In our experience at Rifle, spatial variability in the extractions performed on cored sediments at the Rifle site can be considerable. In this case, the spatial resolution of the sampling scheme should be guided by knowledge gained from the pilot study and cost considerations. We are currently researching in-well sediment incubators as possible surrogates for in situ sediment samples. Initial results suggest that it may be possible to use such incubators to inexpensively characterize both sorbed uranium and in situ microbial response under background and locally biostimulated conditions.

A key piece of information is the amount of U(VI) in groundwater that has been reduced to U(IV) and become associated with the sediments. In this case, preparation and maintenance of reduced uranium in cored sediments for subsequent analysis is a critically important task. A specialized laboratory experiment (Komlos, in prep.) was recently performed where an entire column of sediment was brought intact to a nearby x-ray synchrotron for analysis of the uranium oxidation states in a biostimulated column. When compared to standard handling approaches where sediment was removed from columns and shipped offsite, this approach resulted in considerably higher measurements of reduced uranium in sediments.

4.2 Group 2: Conceptual Model of Uranium Transport and Fate

We take the view that monitoring uranium levels in groundwater is a necessary but not sufficient performance indicator. The expectation of the Group 2 performance indicators is the assessment and confirmation of the conceptual model for uranium transport and fate. System complexity may affect the delivery of amendments to the subsurface as well as long-term effectiveness. This would include bypass from preferential flow paths, spatially variable depletion of bioavailable terminal electron acceptors that are critical to the maintenance of effective uranium bioremediation, changes in the geochemical

environment (e.g., mineral precipitation) that affect uranium availability and reactivity, and the impact of hydrologic events (e.g., elevated water table, intense rainfall/recharge) on the reoxidation of reduced sediments. Evolving reactivity may also occur over longer time scales needed for permeability reduction due to formation of secondary minerals and post-biostimulation re-equilibration to ambient conditions.

It should be mentioned that the microbially-mediated creation of low oxidation-reduction potential (ORP) conditions [e.g., through Fe(II) and sulfide formation] that thermodynamically favor abiotic uranium reduction have been shown to be quite limited kinetically in natural sediments (Jeon, Dempsey et al. 2005). Thus, the general concern with the maintenance of reducing conditions is with regard to preventing reoxidation of bioreduced uranium, not necessarily the continued production of U(IV). However, we also note that post-amendment removal of U(VI) at the Rifle site may result from biosorption of U(VI). The longevity of such immobilization is not yet known but appears to last at least 2 years, depending on abundance of sulfide precipitation during sulfate reduction.

Key monitoring quantities are summarized in Table 2. Aqueous conditions that can be automatically and continuously logged with an in situ sonde include water depth, pH, ORP, alkalinity, specific conductivity, temperature. At the Rifle site, we have four sondes that are continuously operated with bimonthly downloading of this data. An attractive alternative is to set these systems up with telemetry that will allow the data to be downloaded remotely. Water levels can be particularly important to the regional flow field as well as chemical conditions near the water table. Standard multilevel samplers are very useful but it may not be feasible to deploy them in every well. We have supplemented our multilevel samplers with various passive samplers that can be used to economically collect depth-dependent distributions of aqueous and particulate components (Figure 7). In many cases, we can visually identify depth-dependent behavior by examining the sequence of the sampling cells.

Standard aqueous sampling should be directed at the transport of the injectate (e.g., tracer, electron donor, possibly electron acceptors), consumption of electron donors (e.g., ethanol, acetate, lactate) and electron acceptors (e.g., oxygen, nitrate, sulfate), and biostimulation reaction products [carbonate, ammonium, sulfide, reduced metals such as Fe(II), Mn(II)]. Most recently, a technique using tubing and syringes suspended in wells (Spalding and Watson 2006) has been used to provide the most accurate measurements of dissolved gases such as hydrogen, oxygen, carbon dioxide, and nitrogen. The adaptation of this technique to a passive depth-dependent sampling of the water column promises to provide key feedback on bioremediation progress (e.g., hydrogen and carbon dioxide byproduct) and reoxidation (e.g., dissolved oxygen and nitrogen).

Where possible, sediment extractions should be used to identify uranium form [e.g., U(IV), U(VI) as mineral, surface complex], iron form [Fe(II), Fe(III) as mineral and surface complex], and other secondary minerals (e.g., carbonate, sulfide). At the Naturita site, sediments were suspended directly in wells to provide cheaper and easier access to reacted sediments ((Curtis, Fox et al. 2004), see also discussion under Group 1).

4.3 Group 3: Microbiological Conceptual Model

The Group 3 performance indicators are designed to assess and confirm the microbiological conceptual model underlying the engineered bioremediation. Key issues are the ability to differentiate, where necessary, the dominant microbial populations associated with major TEAPs in the bioremediation scheme. Although there is a sequential nature to these TEAPs, there is growing evidence from the field that multiple TEAPs (e.g., iron and sulfate reduction) can be operating simultaneously in the same aguifer sediments, although likely in different microniches. Succession to a new TEAP ostensibly occurs when the preceding TEAP can no longer compete effectively for the electron donor and as a result loses its primacy. This is significant only if the succeeding microorganisms are less effective at uranium bioreduction than their predecessors, which appears to be the case with the acetate-oxidizing microbial consortia at Rifle. Specialized sampling procedures are necessary to identify the microbial community structure and measure the abundance and activity of the dominant microorganisms. Sampling techniques typically are focused on 1) groundwater. 2) sediments, or 3) in-well coupons or sediment incubators. For sampling groundwater, concentration of the planktonic microbial community by filtration is typical. The volume of sample that is filtered depends on the specific microbial analysis. 16S DNA sampling, for example can be done by filtering relatively small volumes (1 to 3 liters), but more sophisticated techniques such as rRNA require larger volumes (~20 liters and more care in sample handling). Sediment sampling is the most challenging since drilling is usually required. Sample volumes again depend on the proposed analysis, ranging from 25 grams to 0.5 kg or more. In-well coupons such as Bio-Sep beads (see below) are attractive since no drilling is involved, but may be problematic if the well-bore environment is not representative of the surrounding formation and groundwater conditions. Recently, efforts have been made to develop an in-well sediment incubator that eliminates this issue by fully occupying part of the well bore with sediment from the site (Peacock, personal communication). This approach makes it possible to track in situ sediment microbial population changes without drilling.

Microbial Analysis Techniques. Phospholipid fatty acid profiles (PLFA) provide a broad but reliable picture of microbial community structure with relatively small groundwater or sediment samples required. Furthermore PLFA analyses are commercially available. Standard molecular biology techniques such as 16-S Clone libraries can also be routinely performed. For some microbial techniques, large volumes (~20 L) of groundwater must be filtered to concentrate sufficient biomass to perform analyses (e.g. gene expression or mRNA). However, molecular biology techniques are evolving rapidly and it is now possible to track both gene expression and mRNA during the course of biostimulation (Holmes, Nevin et al. 2004; Holmes, Nevin et al. 2005). DNA chip arrays and bead arrays make it possible to screen samples for both microbial metabolic function and genetic identity. Many of these tools are used primarily for research, but are quickly becoming available to meet the needs of applied bioremediation. Sediment analyses include the use of similar tools, but require extraction of DNA prior to analysis.

A novel bio-trap technique that captures microbial community structure in the subsurface is Bio-Sep® Beads (Peacock, Chang et al. 2004). The beads are 2-3 mm in diameter, with high porosity and surface area. Biofilms form rapidly in the Bio-Sep® Beads and the biofilm community structure on the beads is more indicative of *in situ* microbial ecology than samples of planktonic organisms. Standard phospholipid fatty acid analysis (PLFA) and DGGE analysis of DNA are performed on extracts from the samplers. The beads serve as an excellent technique for assessing the effects of biostimulation on microbial biomass, community composition, and metabolic state. Microbial analysis of these beads are commercially available.

Fe-reduction and sulfate reduction TEAPs are also amenable to indirect assessment using geophysical monitoring via electrical methods (Williams, Kemna et al. 2006). Recent unpublished results from the Rifle site using electrical resistivity tomography indicate that iron reducing and sulfate reducing conditions are readily distinguished in adjacent electron donor amendment plots one of which was driven to sulfate reduction and the other maintained in Fe reduction. These results are preliminary but indicate the likely value of electrical methods for minimally invasive means of assessing microbial processes in the subsurface.

5 Summary and Prioritization of Performance Indicators

Table 2 summarizes the performance indicators. Priorities are designated in three categories, mandatory, desirable, and optional. In addition, the temporal dimension of the monitoring regime is provided as a guideline for monitoring frequency. Mandatory performance indicators are those indicators judged to be the minimum set of data needed to validate a uranium bioremediation project. Desirable performance indicators are those, which, if included, could be used to make a stronger case for the success and effectiveness of uranium bioremediation. Optional performance indicators are the lowest priority, but may provide valuable information in selected cases or enhance the understanding of system processes. This prioritization scheme was developed to minimize the cost of system assessment by focusing on low cost measurements in the mandatory category while still ensuring that critical data are obtained. Site specific conceptual or numerical models may indicate different priorities or the need for emphasis on specific measurements.

It is recognized that uranium contamination in groundwater occurs in a variety of hydrogeologic settings and that some settings may be amenable to different priorities. For example, reclamation and clean up of sites where uranium has been mined by In Situ Leaching (ISL) may have a different set of issues and drivers than alluvial aquifers contaminated during uranium milling operations. ISL site are commonly deeper than milling tailings sites, but have the advantage that existing wells and infrastructure can be used for cleanup and monitoring using geophysical techniques. ISL cleanup is commonly done abiotically by sweeping clean water through the system to dilute uranium concentrations followed by reduction by H₂S and additional pumping to remove residuals. Bioremediation could decrease cost and increase the effectiveness of

treatment. Geophysical and biogeochemical monitoring is likely critical in such cases to demonstrate effective spatial distribution of treatment and achievement of desired results over appropriate time periods.

The most complete set of performance indicators will not be useful unless effectively analyzed and communicated to problem holders, regulators, and stakeholders. Visualization of the spatial and temporal changes in performance indicators is a particularly useful analysis and communication tool. Such visualization can be as simple as performance indicator versus time plots posted on a borehole map. Figure 9 shows the concentration of U(VI) in space for the Rifle site. A series of such plots are used to animate the visual evidence for U(VI) removal over a 3 month period for the 2002 Rifle field experiment. The animation illustrates key points about the change in U(VI) reduction at the onset of sulfate reduction.

Performance indicators are typically also used to update numerical modeling of the system. For example, if groundwater elevations indicate that permeability of the treatment zone is changing, different values of permeability can be input to the numerical model to assess the extent of change that is consistent with water table elevations. The updated model can then be used to assess the effectiveness of treatment and degree of rerouting of groundwater flow that may be occurring. Such assessments can also be used to estimate the value of additional monitoring points or to indicate that some monitoring point may no longer be cost effective to sample.

Acknowledgments

Preparation of this document was funded by the U.S. Nuclear Regulatory Commission, Research Division, Thomas Nicholson, Manager. Our research in uranium bioremediation at the Rifle site is funded by the U.S. Department of Energy, Office of Science, Biological and Environmental Research, Environmental and Remediation Sciences Program. Pacific Northwest National Laboratory is operated by Battelle Memorial Institute for the U.S. Department of Energy under contract DE-AC06-76RLO 1830.

References Cited

- Abdelouas, A., W. Lutze, et al. (1999). "Oxidative dissolution of uraninite precipitated on Navajo sandstone." <u>Journal of Contaminant Hydrology</u> **36**(3-4): 353-375.
- Anderson, R. T., H. A. Vrionis, et al. (2003). "Stimulating the in situ activity of Geobacter species to remove uranium from the groundwater of a uranium-contaminated aquifer." <u>Applied and Environmental Microbiology</u> **69**(10): 5884-5891.
- Arai, Y., M. McBeath, et al. (2006). "Uranyl adsorption and surface speciation at the imogolite-water interface: Self-consistent spectroscopic and surface complexation models." <u>Geochimica Et Cosmochimica Acta</u> **70**(10): 2492-2509.
- Burns, P. C. (1999). The Crystal Chemistry of Uranium. <u>Uranium: Mineralogy, Geochemistry, and the Environment</u>. P. C. Burns and R. Finch. Washington, D.C., Mineralogical Society of America. **38:** 23-90.
- Curtis, G. P., P. Fox, et al. (2004). "Comparison of in situ uranium K-D values with a laboratory determined surface complexation model." <u>Applied Geochemistry</u> **19**(10): 1643-1653.
- Davis, J. A. and D. B. Kent (1990). "Surface Complexation Modeling in Aqueous Geochemistry." <u>Reviews in Mineralogy</u> **23**: 177-260.
- DOE (1999). Final site observational work plan for the UMTRA project Old Rifle site GJO-99-88-TAR. Grand Junction, Colo.
- Gee, G. W., A. L. Ward, et al. (2002). "A vadose zone water fluxmeter with divergence control." <u>Water Resources Research</u> **38**(8): -.
- Holmes, D. E., K. P. Nevin, et al. (2004). "In situ expression of nifD in Geobacteraceae in subsurface sediments." <u>Applied and Environmental Microbiology</u> **70**(12): 7251-7259.
- Holmes, D. E., K. P. Nevin, et al. (2005). "Potential for quantifying expression of the Geobacteraceae citrate synthase gene to assess the activity of Geobacteraceae in the subsurface and on current-harvesting electrodes." <u>Applied and Environmental Microbiology</u> **71**(11): 6870-6877.
- Jeon, B. H., B. A. Dempsey, et al. (2005). "Chemical reduction of U(VI) by Fe(II) at the solid-water interface using natural and synthetic Fe(III) oxides." <u>Environmental Science & Technology</u> **39**(15): 5642-5649.
- LaBolle, E. M. and G. E. Fogg (2001). "Role of molecular diffusion in contaminant migration and recovery in an alluvial aquifer system." <u>Transport in Porous Media</u> **42**(1-2): 155-179.

- Lovley, D. R., E. J. P. Phillips, et al. (1991). "Microbial Reduction of Uranium." <u>Nature</u> **350**(6317): 413-416.
- Payne, T. E., J. A. Davis, et al. (1996). "Uranium adsorption on ferrihydrite Effects of phosphate and humic acid." <u>Radiochimica Acta</u> **74**: 239-243.
- Peacock, A. D., Y. J. Chang, et al. (2004). "Utilization of microbial biofilms as monitors of bioremediation." <u>Microbial Ecology</u> **47**(3): 284-292.
- Qafoku, N. P., J. M. Zachara, et al. (2005). "Kinetic desorption and sorption of U(VI) during reactive transport in a contaminated Hanford sediment." <u>Environmental Science & Technology</u> **39**(9): 3157-3165.
- Roden, E. E. and T. D. Scheibe (2005). "Conceptual and numerical model of uranium(VI) reductive immobilization in fractured subsurface sediments." <u>Chemosphere</u> **59**(5): 617-628.
- Scheibe, T. D., Y. Fang, et al. (2006). "Transport and biogeochemical reactions of metals in a chemically heterogeneous aquifer." <u>Geosphere</u> in press.
- Suzuki, Y., S. D. Kelly, et al. (2002). "Radionuclide contamination Nanometre-size products of uranium bioreduction." Nature **419**(6903): 134-134.
- Waite, T. D., J. A. Davis, et al. (1994). "Uranium(Vi) Adsorption to Ferrihydrite Application of a Surface Complexation Model." <u>Geochimica Et Cosmochimica Acta</u> **58**(24): 5465-5478.
- Williams, K. H., A. Kemna, et al. (2006). The Role of Geoelectrical Methods in Monitoring Stimulated Sulfate-Reduction: Insights Gained From Field-Scale Experiments. <u>AGU Fall Meet. Suppl.</u>, Eos Trans. 87(52) Abstract NS21A-06

Tables

Table 1. Environmentally important redox reactions in order of decreasing energy yield.

Environmentally Important Redox Reactions

Reaction	E _h (V)	ΔG
Reduction of O ₂		
O ₂ + 4H ⁺ +4e ⁻ > 2H ₂ O	+0.812	-29.9
Reduction of NO₃⁻		
$2NO_3^- + 6H^+ + 6e^> N_2 + 3H_2O$	+0.747	-28.4
Reduction of Mn ⁴⁺		
$MnO_2 + 4H^+ + 2e^> Mn^{2+} + 2H_2O$	+0.526	-23.3
Reduction of Fe ³⁺		
$Fe(OH)_3 + 3H^+ + e^> Fe^{2+} + 3H_2O$	-0.047	-10.1
Reduction of SO ₄ ² -		
SO ₄ ²⁻ + 10H ⁺ + 8e ⁻ > H ₂ S + 4H ₂ O	-0.221	-5.9
Reduction of CO ₂		
CO ₂ + 8H ⁺ + 8e ⁻ > CH ₄ + 2H ₂ O	-0.244	-5.6



Table 2. Prioritized Information and Monitoring Parameters for Assessment of Bioremediation of U(VI) in Groundwater

Mandatory Uranium Distribution, Magnitude, Form, and Mobility

Information area/parameter	Desired Range	Comments
Site conceptual model for uranium source term	NA	Consideration of alternative conceptual models critical
Spatial extent of contamination zone (plume geometry)	±20% of estimate	Differientiate between vadose zone and aquifer concentrations; aqueous and sediment associated uranium; geometry drive layout of bioremediation system
Form and mobility/lability	± 30% of estimate	Mobility based on uranium form, potential for future mobility based on labile fraction

Mandatory Hydrologic and Geologic Site Information

Information area/parameter	Desired Range	Comments
Site conceptual model for		
subsurface (vadose zone and		
groundwater) flow and		Consideration of alternative conceptual models
contaminant transport	NA	critical
Temporal recharge	±20%	Seasonal and episodic impact to unsaturated flow, extreme recharge event and impact must be considered if flooding probable at the site
Vadose zone hydrogeology:		Ĭ.
porosity, water retention	±20% of	Seasonal and episodic impact to flow direction
function parameters	estimate	critical
Groundwater flow velocity	±30% of	
(Darcy flux) and direction	estimate	Seasonal and episodic impact critical
Water table dynamics (use hourly data as event-based geochemical sampling driver)	NA	Relationship between water table and U concentration critical
Site hydrogeology:		
hydraulic conductivity,		Fundamental to both site and process conceptual
porosity, and dispersivity	NA	model
Remediation process	NI A	Fundamental to prioritization of monitoring
conceptual model	NA NA	parameters

Particle size characteristics NA Reactive surface area, clays, upscaling lab to field

Mandatory Geochemical and Microbiological Monitoring Parameters

Parameter Parameter	Desired Range	Comments
Background U(VI)	Desired Range	Comments
concentration, monthly or		
bi-monthly and event-base		Number of samping points based on plume and
(e.g. high water table)	NA	treatment zone complexity and size
Treatment zone U(VI)		Number of sampling points based on plume and
concentration	Below MCL	treatment zone complexity and size
DO, ORP, specific	DO<0.5,	, , , , , , , , , , , , , , , , , , ,
conductivity, and pH	ORP<0,	
measured hourly to 4 times	conducitivity	
daily in background and	initial	
treatment zone (autonomous	increase, pH	Values used as overall dynamic indicator of impact
multiparameter probes)	~ steady	of bioremediation on subsurface geochemistry
DO, ORP, specific		
conductivity, and pH		
measured at time of		
groundwater sampling in		
background and treatment		Linkage of U(VI) concentrations with parameter
zone using flow-cell with		change evidence for bioremediation process
multiparameter probe	See above	conceptual model
Aqueous electron acceptors		
and reduction byproducts in		
background and treatment		Significant concentrations of oxygen and/or other
zone: nitrate, nitrite,		electron acceptors above the U TEAP on the redox
ammonium, Mn(IV/II),		ladder must be addressed by the bioremediation
sulfate, sulfide	NA	strategy and their reduction products monitored
		Fe(III) minerals provide sorption sites for Fe(II) &
		U(VI), terminal electron acceptor for Fe-reducing
Fe(III) mineral abundance	NA	bacteria, dissolved Fe(II) source
	Increasing	
	Fe(II); sulfide	Maintaining metal reduction may optimize U(VI)
Fe(II), sulfide measured in	indicator of	removal from groundwater; sulfate reduction may
field at time of sampling for	sulfate	enhance long-term immobilization in sulfate-rich
U(VI)	reduction	systems
		Evidence of delivery and treatment zone
Electron donor concentration		distribution; consumption calculation based on
in treatment zone	>0	tracer data
	>0 in	
	treatment	Typically Br is used for conservative tracer,
Tracer for electron donor	zone	accurate indication of donor distribution
Alkalinity (measured in the		
field)	NA	Indicator of carbonate geochemistry

Desirable Monitoring Parameters

Desirable Montelling Farameters			
Parameter	Desired Range	Comments	
Depth discrete U(VI) data (upper/mid/lower part of contaminated zone)	MCL	Decreased effectiveness of treatment in the uppermost part of the saturated zone may be problematic	
Major cations and anions	NA	Provides additional evidence for dominant geochemical reactions	
Impact of treatment process on groundwater flow directions (hourly water level at minimum 4 points)	Dependent on background flow	Helps to provide assurance that groundwater is not rerouted around treatment zone	
In situ redox status of U using in situ sediment incubators (ISI's)	Significant U(IV) present	Evidence that precipitation of U(IV) is occurring in situ	
Microbiological assessment using coupons or in situ incubators	Shift to metal and/or sulfate reduction	Evidence for desired in situ microbial respiration obtained from deploying coupons or in situ incubators in well bores and periodically measuring microbial parameters (see text for additional discussion)	
Depth-discrete sediment sampling/extraction for U, Fe, AVS	NA	Evidence for conversion of terminal electron acceptors	
Major dissolved gas components in groundwater	NA	Evidence for key TEAPs and microbial metabolism	
GPR Cross-well Measurements	Shift in radar velocity in zone of electron donor	Defines 2-D distribution of electron donor	

Optional Monitoring Parameters

Optional Montelling Larameters			
Parameter	Desired Range	Comments	
Depth-discrete data for mandatory geochemical parameters	NA	Characterizes spatial distribution of fundamental biogeochemistry in aquifer	
Depth-discrete data for desirable monitoring parameters	NA	Characterizes spatial distribution of desired biogeochemical reactions in aquifer	
Impact of treatment process on hydraulic properties	<15% change	Documents possible system clogging of pores	
Organic and inorganic carbon analyses	NA	More accurate documentation of carbonate geochemistry	
Microbiological assessment performed directly on sampling of treatment zone materials	Shift to metal and/or sulfate reduction	Measurements directly on groundwater filtrates or sediment cores provide "gold standard" assessment of microbial community structure (e.g PLFA, 16S, DNA/RNA chip arrays, or functional chip arrays)	
In situ redox status of U by direct sampling of in situ materials		U(IV)/U(VI) measurements on in situ sediments provide "ground truth" for U bioreduction	
Electrical Resistivity and Self Potential Tomography	NA	Defines 3-D arrangement of dominant TEAP's	
Seismic reflection	NA	Sensitive to gas evolution and secondary mineral precipitation	

Temporal Dimension of Monitoring Regime (see also flow chart in text)

Information area/parameter	Desired Range	Comments
1st Year: monthly or bi- monthly sampling	NA	Sampling during early stage of treatment increases chance of capturing metal reduction phase
2nd and 3rd Year: bi- monthly sampling	NA	Adjustments to sampling frequency needed based on expected life of electron donor or other facets of treatment system
3rd through 10th year: quarterly sampling conditioned on water table behavior	NA	Seasonal and water table fluctuations should drive sampling schedule
Beyond 10 years, adaptive sampling based on prior year results	NA	If a high level of confidence can be demonstrated based on sampling over fewer than 10 years, adaptive sampling strategies could start earlier

Figures

Figure 1. Dissolved uranium as a function of pH, carbonate, nitrate and sulfate (Morrison et al. 1995)

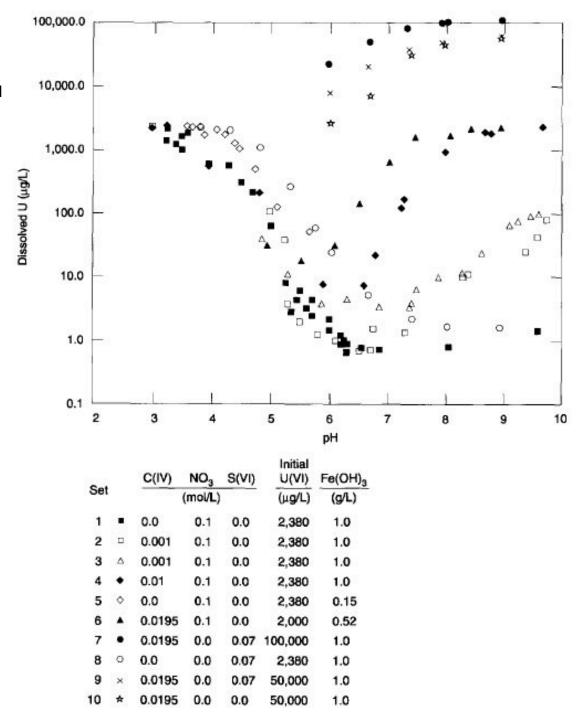
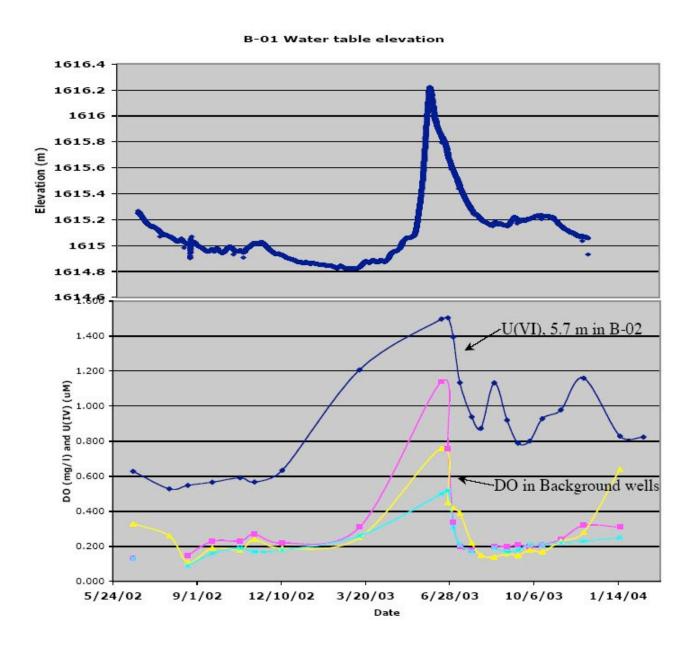


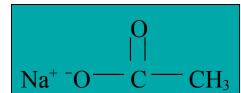
Figure 2. Changes in dissolved oxygen and U(VI) concentration with water table rise.



DO (mg/l) 1.5 0.5 1 2 2.5 3 Figure 3. Oxygen stratification observations and modeling. 0 A. Examples of DO stratification. B. Four model cases showing that microbial consumption of oxygen is needed M-02 2003 M-16 2005 to match observations. 2 Ê 3 Depth 4 B. 2.0E-05 6.0E-05 1.0E-04 1.4E-04 1.8E-04 2.2E-04 2.0E-05 6.0E-05 1.0E-04 1.4E-04 1.8E-04 2.2E-04 40 30 30 N 20 N 20 O₂ influent with GW O₂ microbial TEAP Flow 0 50 100 150 200 250 300 50 100 150 200 250 300 40 30 + O₂ microbial TEAP N 20 **N** 20 O₂ diffusion at WT Over time 10 150 200 100 250 300 50 100 250 50 200 150 300

Fig. 4. Microbial mediation of Fe(III) Reduction. U(VI) is the mobile valence state of uranium whereas reduced uranium, U(IV), is insoluble as uraninite. Reduction of U(VI) to U(IV) within aquifers precipitates and immobilizes uranium per lab studies that suggest a simple strategy to promote U(VI) reduction in contaminated aquifers is to add acetate as an electron donor to stimulate dissimilatory metal-reducing microorganisms. U(VI) is reduced concurrently with Fe(III). Original concept from Lovley et al. 1991. Field implementation by Anderson et al 2003.

Acetate structure



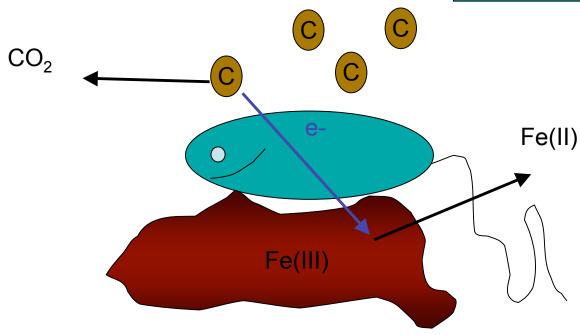
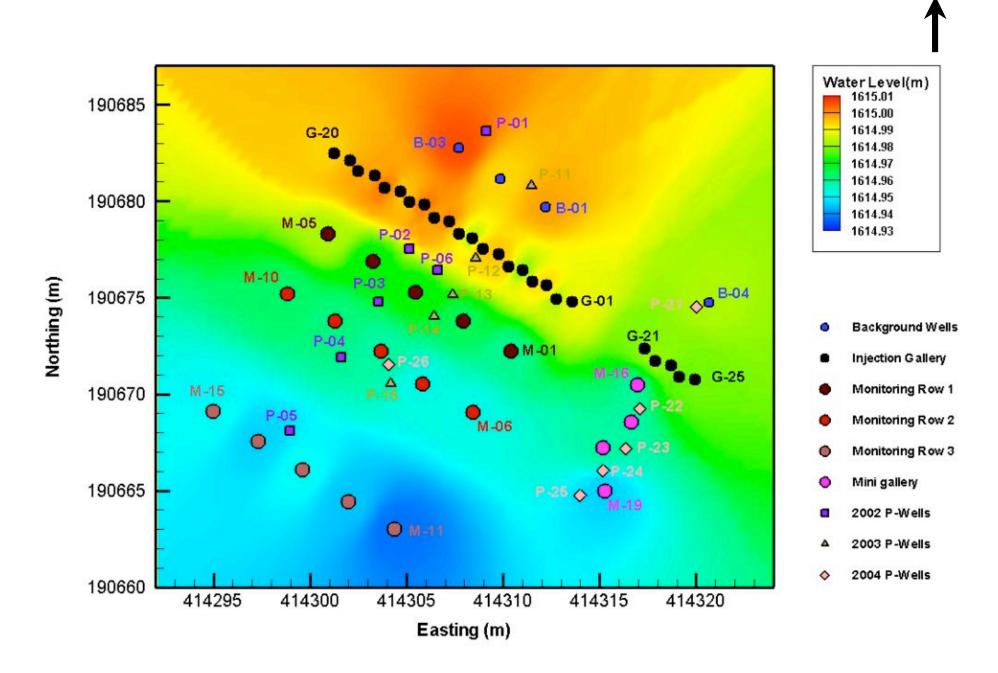


Fig. 5 Borehole arrays for pilot-scale testing at the Rifle site. A. Layout of wells showing water table elevations. general groundwater flow direction to the southwest.

N



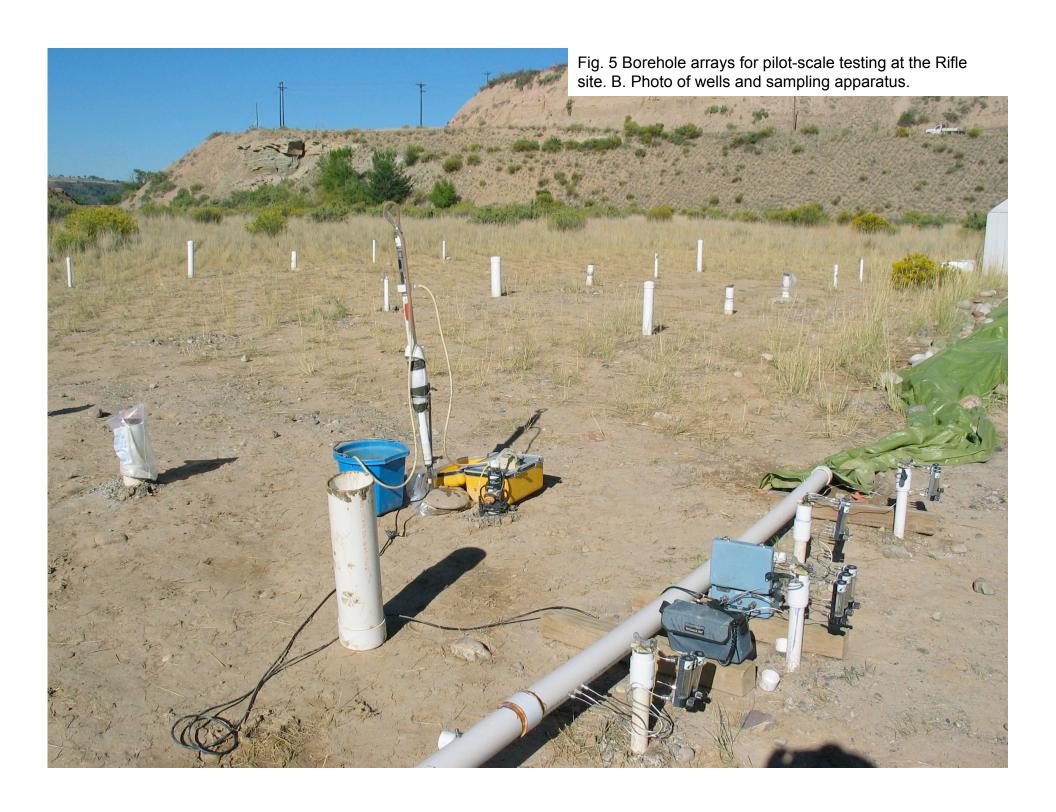


Figure 6. Distribution of U(VI) as a function of depth during a biostimulation experiment. Note increase in U(VI) concentration near water table on 7/30/2002. Data obtained using passive multilevel samplers.

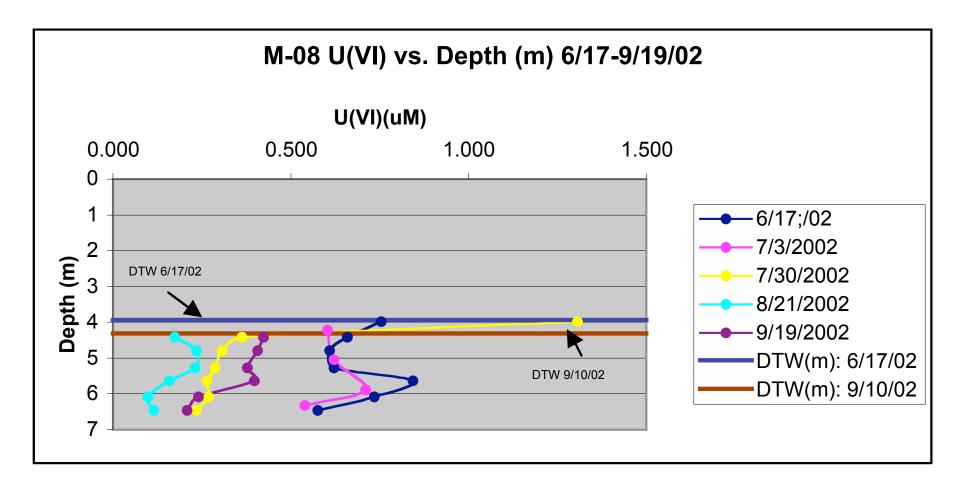
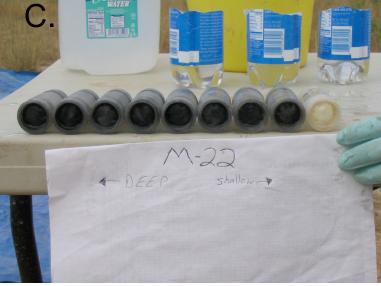
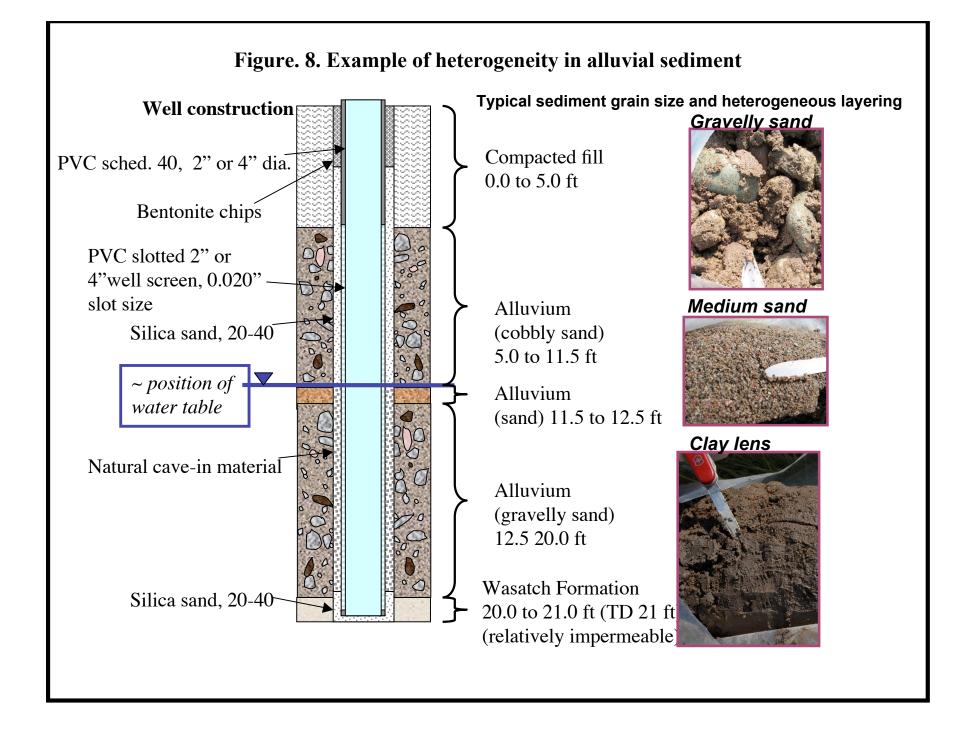


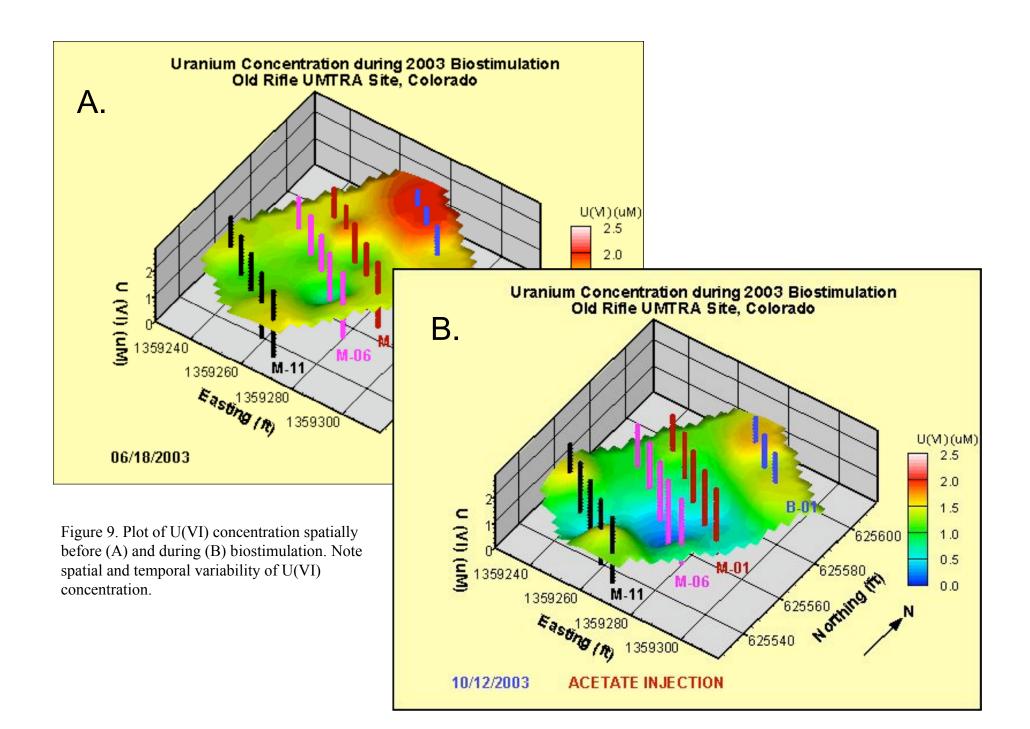
Figure 7. Passive multilevel samplers. A. Cell on support rod being lowered into monitoring well. B. MLS cells from a background well. C. MLS cells from a treatment zone well undergoing sulfate reduction.











Bibliography

- Abdel Aal, G. Z., E. A. Atekwana, et al. (2004). "Effects of microbial processes on electrolytic and interfacial electrical properties of unconsolidated sediments." <u>Geophysical Research Letters</u> **31**(12): L12505.
- Abdelouas, A., W. Lutze, et al. (1999). "Oxidative dissolution of uraninite precipitated on Navajo sandstone." <u>Journal of Contaminant Hydrology</u> **36**(3-4): 353-375.
- Anderson, R. T., H. A. Vrionis, et al. (2003). "Stimulating the in situ activity of Geobacter species to remove uranium from the groundwater of a uranium-contaminated aquifer." Applied and Environmental Microbiology **69**(10): 5884-5891.
- Arai, Y., M. Marcus, et al. (2006). Spectroscopic evidence for uranium bearing particulates in vadose zone sediments at Hanford DOE Site, 300-area, Washington. <u>Abstracts</u>, 18th World Congress of Soil Science. Philadelphia, PA.
- Arai, Y., M. McBeath, et al. (2006). "Uranyl adsorption and surface speciation at the imogolite-water interface: Self-consistent spectroscopic and surface complexation models." <u>Geochimica Et Cosmochimica Acta</u> **70**(10): 2492-2509.
- Atekwana, E. A., D. D. W. Jr, et al. (2004). "In-situ apparent conductivity measurements and microbial population distribution at a hydrocarbon-contaminated site." <u>Geophysics</u> **69**(1): 56-63.
- Banfield, J. F., V. N.C., et al. (2005). "Proteogenomic approaches for the molecular characterization of natural microbial communities." <u>OMICS</u> **9**: 301-333.
- Banfield, J. F., S. A. Welch, et al. (2000). "Aggregation-based crystal growth and microstructure development in natural iron oxyhydroxide biomineralization products." <u>Science</u> **289**(5480): 751-754.
- Bernhard, G., G. Geipel, et al. (2001). "Uranyl(VI) carbonate complex formation: Validation of the Ca2UO2(CO3)(3)(aq.) species "Radiochim. Acta 89: 511-518.
- Bond, D. L., J. A. Davis, et al. (2006). "U(VI) release from contaminated vadose zone sediments: Estimation of potential contributions from dissolution and desorption." Geochimica et Cosmochimica Acta **submitted for publication**.
- Brown, D. G., J. Komlos, et al. (2005). "Simultaneous utilization of acetate and hydrogen by Geobacter sulfurreducens and implications for use of hydrogen as an indicator of redox conditions." <u>Environmental Science & Technology</u> **39**(9): 3069-3076.
- Brown, S. D., M. R. Thompson, et al. (2006). "Molecular dynamics of the Shewanella oneidensis response to chromate stress." <u>Molecular & Cellular Proteomics</u> **5**(6): 1054-1071.
- Buck, E. C., N. R. Brown, et al. (1996). "Contaminant uranium phases and leaching at the Fernald site in Ohio." <u>Environmental Science & Technology</u> **30**(1): 81-88.
- Burns, P. C. (1999). The Crystal Chemistry of Uranium. <u>Uranium: Mineralogy, Geochemistry, and the Environment</u>. P. C. Burns and R. Finch. Washington, D.C., Mineralogical Society of America. **38:** 23-90.
- Callister, S. J., R. C. Barry, et al. (2006). "Normalization approaches for removing systematic biases associated with mass spectrometry and label-free proteomics." <u>Journal of Proteome Research</u> **5**(2): 277-286.
- Callister, S. J., C. D. Goddard, et al. (2006). "Comparison of aerobic and photosynthetic

- Rhodobacter sphaeroides proteomes." JPR in press.
- Catalano, J. G. and G. E. Brown (2005). "Uranyl adsorption onto montmorillonite: Evaluation of binding sites and carbonate complexation." <u>Geochimica Et Cosmochimica Acta</u> **69**(12): 2995-3005.
- Catalano, J. G., J. P. McKinley, et al. (2006). "Changes in uranium speciation through a depth sequence of contaminated Hanford sediments." <u>Environmental Science & Technology</u> **40**(8): 2517-2524.
- Chan, C. S., G. De Stasio, et al. (2004). "Microbial polysaccharides template assembly of nanocrystal fibers." <u>Science</u> **303**(5664): 1656-1658.
- Chandler, D. P., A. E. Jarrell, et al. (2006). "Suspension array analysis of 16S rRNA from Fe- and $SO_4^{2^-}$ reducing bacteria in uranium contaminated sediments undergoing bioremediation." <u>Appl. Environ. Microbiol.</u> **Submitted**.
- Chang, Y. J., P. E. Long, et al. (2005). "Microbial incorporation of C-13-labeled acetate at the field scale: Detection of microbes responsible for reduction of U(VI)." Environmental Science & Technology **39**(23): 9039-9048.
- Chapelle, F. H., S. K. Haack, et al. (1996). "Comparison of E(h) and H-2 measurements for delineating redox processes in a contaminated aquifer." <u>Environmental Science & Technology</u> **30**(12): 3565-3569.
- Chapelle, F. H., P. B. Mcmahon, et al. (1995). "Deducing the Distribution of Terminal Electron-Accepting Processes in Hydrologically Diverse Groundwater Systems." Water Resources Research **31**(2): 359-371.
- Chen, J., S. Hubbard, et al. (2006). "Development of a joint hydrogeophysical inversion approach and application to a contaminated fractured aquifer." <u>Water Resources Research</u> in press.
- Chen, J., S. Hubbard, et al. (2004). "Geochemical characterization using geophysical data and Markov Chain Monte Carlo methods: A case study at the South Oyster Bacterial transport site in Virginia." <u>Water Resources Research</u> **40**(W12412): doi: 1029/2003WR002883.
- Choi, J., D. Meldrum, et al. (2006). Development of cellular absorptive tracers (CATs) for a quantitative characterization of the complexity of nanoscale biological systems. <u>232nd ACS National Meeting</u>. San Francisco, CA.
- Corredor, J. E., B. Wawrik, et al. (2004). "Geochemical Rate-RNA integration study: Ribulose-1,5-bisphosphate carboxylase/oxygenase gene transcription and photosynthetic capacity of planktonic photoautotrophs." Applied and Environmental Microbiology **70**(9): 5559-5568.
- Curtis, G. P., J. A. Davis, et al. (2006). "Simulation of reactive transport of uranium(VI) in groundwater with variable chemical conditions." <u>Water Resources Research</u> **42**(4): -.
- Curtis, G. P., P. Fox, et al. (2004). "Comparison of in situ uranium K-D values with a laboratory determined surface complexation model." <u>Applied Geochemistry</u> **19**(10): 1643-1653.
- Davis, J. A., G. P. Curtis, et al. (2006). "Processes affecting transport of uranium in a suboxic aquifer." Physics and Chemistry of the Earth in press.
- Davis, J. A. and D. B. Kent (1990). "Surface Complexation Modeling in Aqueous Geochemistry." Reviews in Mineralogy **23**: 177-260.
- Davis, J. A., D. E. Meece, et al. (2004). "Approaches to surface complexation modeling

- of uranium(VI) adsorption on aquifer sediments." <u>Geochimica Et Cosmochimica Acta</u> **68**(18): 3621-3641.
- Ding, Y., K. Hixson, et al. (2006). "The proteome of dissimilatory metal-reducing microorganism Geobacter sulfurreducens under various growth conditions." <u>Biochim Biophys Acta</u> **Epub ahead of print**.
- DOE (1999). Final site observational work plan for the UMTRA project Old Rifle site GJO-99-88-TAR. Grand Junction, Colo.
- DOE (2006). Environmental remediation sciences program strategic plan, Version 9, Office of Biological and Environmental Reseach, Environmental Remediation Sciences Division.
- Dong, H. L., J. E. Kostka, et al. (2003). "Microscopic evidence for microbial dissolution of smectite." <u>Clays and Clay Minerals</u> **51**(5): 502-512.
- Dowling, N. J. E., F. Widdel, et al. (1986). "Phospholipid ester-linked fatty acid biomarkers of acetate-oxidizing sulfate reducers and other sulfide forming bacteria. ." J. Gen. Microbiol. **132**: 1815-1825.
- Elias, D. A., J. M. Senko, et al. (2003). "A procedure for quantitation of total oxidized uranium for bioremediation studies." <u>Journal of Microbiological Methods</u> **53**(3): 343-353.
- Energy, U. S. D. o. (1999). Final Site Observational Work Plan for the UMTRA Project Old Rifle Site. DOE.
- Fang, R., D. Elias, et al. (2006). "Differential label-free quantitative proteomic analysis of Shewanella oneidensis cultured under aerobic and suboxic conditions by accurate mass and time tag approach." Mol Cell Proteomics **5**(4): 714-725.
- Fang, Y. L., S. B. Yabusaki, et al. (2006). "A general simulator for reaction-based biogeochemical processes." Computers & Geosciences **32**(1): 64-72.
- Fang, Y. L., G. T. Yeh, et al. (2003). "A general paradigm to model reaction-based biogeochemical processes in batch systems." Water Resources Research **39**(4):
- Fazio, S. A., D. J. Uhlinger, et al. (1982). "Estimations of uronic acids as quantitative measures of extracellular and cell wall polysaccharide polymers from environmental samples." <u>Applied and Environmental Microbiology</u> **43**: 1151-1159.
- Finneran, K. T., R. T. Anderson, et al. (2002). "Potential for Bioremediation of uranium-contaminated aquifers with microbial U(VI) reduction." <u>Soil & Sediment</u> Contamination **11**(3): 339-357.
- Gallagher, E., L. McGuinness, et al. (2005). "13C-carrier DNA shortens the incubation time needed to detect benzoate-utilizing denitrifying bacteria by stable-isotope probing." <u>Applied and Environmental Microbiology</u> **71**(9): 5192-5196.
- Gee, G. W., A. L. Ward, et al. (2002). "A vadose zone water fluxmeter with divergence control." <u>Water Resources Research</u> **38**(8): -.
- Gelhar, L. W. (1993). <u>Stochastic Subsurface Hydrology</u>. New Jersey, Prentice Hall.
- Gorby, Y. A. and D. R. Lovley (1992). "Enzymatic Uranium Precipitation." <u>Environmental Science & Technology</u> **26**(1): 205-207.
- Guckert, J. B., M. A. Hood, et al. (1986). "Phospholipid, ester-linked fatty acid profile changes during nutrient deprivation of Vibrio cholerae: increases in the trans/cis ratio and proportions of cyclopropyl fatty acids." <u>Appl. Environ. Microbiol.</u> **52**: 794-801.

- Guillaumont, R., T. Fanghanel, et al. (2003). Chemical Thermodynamics 5. Update on the Chemical Thermodynamics of Uranium, Neptunium, Plutonium, Americium, and Technetium. Nuclear Energy Agency. Amsterdam, Elsevier.
- Hazen, T. C., D. Joyner, et al. (2004). Functional Microbial Changes During Lactate and HRC-Stimulated Bioreduction of Cr(VI) in Hanford 100H Sediments. <u>American Society for Microbiology 104th Meeting</u>. New Orleans, LA.
- Hegde, P. S., I. R. White, et al. (2003). "Interplay of transcriptomics and proteomics." <u>Current Opinion in Biotechnology</u> **14**: 647-651.
- Holmes, D., R. O'Neil, et al. (2005). Gene Expression Analysis of *in situ* Metabolism and Nutrient Status of Geobacteraceae in Subsurface Environments. <u>The 105th</u> General Meeting of the American Society for Microbiology. Atlanta, GA.
- Holmes, D. E., K. P. Nevin, et al. (2004). "In situ expression of nifD in Geobacteraceae in subsurface sediments." <u>Applied and Environmental Microbiology</u> **70**(12): 7251-7259.
- Holmes, D. E., K. P. Nevin, et al. (2005). "Potential for quantifying expression of the Geobacteraceae citrate synthase gene to assess the activity of Geobacteraceae in the subsurface and on current-harvesting electrodes." <u>Applied and Environmental Microbiology</u> **71**(11): 6870-6877.
- Hubbard, S. S., J. Chen, et al. (2001). "Hydrogeological characterization of the South Oyster Bacterial Transport site using geophysical data." <u>Water Resources Research</u> **37**(10): 2431-2456.
- Hubbard, S. S., J. S. Chen, et al. (2001). "Hydrogeological characterization of the South Oyster Bacterial Transport Site using geophysical data." <u>Water Resources Research</u> **37**(10): 2431-2456.
- Hubbard, S. S., J. Peterson, et al. (1997). "Estimation of permeable pathways and water content using tomographic radar data." <u>The Leading Edge</u> **16**(11): 1623-1628.
- Hyndman, D. W. and S. Gorelick (1996). "Estimating lithologic and transport properties in three dimensions using seismic and tracer data: The Kesterson aquifer." <u>Water Resources Research</u> **32**(9): 2659-2670.
- Jaffé, P. R., J. Komlos, et al. (2006). Reduction and Re-oxidation of Soils During and After Uranium Bioremediation; Implications for Long Term Uraninite Stability and Bioremediation Scheme Implementation (PI meeting poster presentation) ERSP_Meeting. Airlie Center, Warrenton, VA.
- Jaisi, D. P., R. K. Kukkadapu, et al. (2005). "Control of Fe(III) site occupancy on the rate and extent of microbial reduction of Fe(III) in nontronite." <u>Geochimica Et Cosmochimica Acta</u> **69**(23): 5429-5440.
- Jeon, B. H., B. A. Dempsey, et al. (2005). "Chemical reduction of U(VI) by Fe(II) at the solid-water interface using natural and synthetic Fe(III) oxides." Environmental Science & Technology **39**(15): 5642-5649.
- Jeon, B. H., B. A. Dempsey, et al. (2003). "Sorption kinetics of Fe(II), Zn(II), Co(II), Ni(II), Cd(II), and Fe(II)/Me(II) onto hematite." Water Research 37(17): 4135-4142.
- Jeon, B. H., S. D. Kelly, et al. (2004). "Microbial reduction of U(VI) at the solid-water interface." Environmental Science & Technology **38**(21): 5649-5655.
- Jin, Q. S. and C. M. Bethke (2005). "Predicting the rate of microbial respiration in geochemical environments." <u>Geochimica Et Cosmochimica Acta</u> **69**(5): 1133-

- Kalmykov, S. N. and G. R. Choppin (2000). "Mixed Ca2+/UO22+/CO32- complex formation at different ionic strengths." Radiochimica Acta **88**(9-11): 603-606.
- Kemna, A., A. Binley, et al. (2000). "Complex resistivity tomography for environmental applications." <u>Chemical Engineering Journal</u> **77**: 11-18.
- Kemna, A., A. Binley, et al. (2004). "Crosshole IP imaging for engineering and environmental applications." <u>Geophysics</u> **69**(1): 97-107.
- Kerkhof, L., L. McGuinness, et al. (2006). Acetate utilizing bacteria in groundwater and on soil at the Rifle, CO UMTRA site based on stable isotope probing (SIP) (in prep.). EMSP PI meeting. Warrenton, Virginia.
- Kerkhof, L., M. Santoro, et al. (2000). "Response of Soybean Rhizosphere Communities to Human Hygiene Water Addition as Determined by Community-Level Physiological Profiling (CLPP) and Terminal Restriction Fragment Length Polymorphism (TRFLP)." FEMS Microbiology Letters 184: 95-101.
- Kerkhof, L. and B. B. Ward (1993). "Comparison of Nucleic-Acid Hybridization and Fluorometry for Measurement of the Relationship between Rna/DNA Ratio and Growth-Rate in a Marine Bacterium." <u>Applied and Environmental Microbiology</u> **59**(5): 1303-1309.
- Kindred, J. S. and M. A. Celia (1989). "Contaminant Transport and Biodegradation .2. Conceptual-Model and Test Simulations." <u>Water Resources Research</u> **25**(6): 1149-1159.
- Kitano, H. (2002). "Computational systems biology." Nature **420**(6912): 206-210.
- Kitano, H. (2002). "Systems biology: A brief overview." Science 295(5560): 1662-1664.
- Kohler, M., G. P. Curtis, et al. (2004). "Methods for estimating adsorbed uranium(VI) and distribution coefficients of contaminated sediments." Environmental Science & Technology **38**(1): 240-247.
- Komlos, J. and P. R. Jaffe (2004). "Effect of iron bioavailability on dissolved hydrogen concentrations during microbial iron reduction." <u>Biodegradation</u> **15**(5): 315-325.
- Komlos J., K. R. K., Zachara J.M., Jaffé P.R. (2006). <u>Re-Oxidation of Uranium Precipitated During Long-Term Iron/Uranium Reducing Conditions</u>. American Chemical Society 232nd National Meeting & Exposition.
- Konopka, A. (2004). "Systems biology not brand new, but different." <u>ASM News</u> **70**(4): 163-168.
- Konstantinidis, K. T. and J. M. Tiedje (2005). "Genomic insights enhance the species definition for prokaryotes." PNAS 102: 2567-2572.
- Kostka, J. E., D. D. Dalton, et al. (2002). "Growth of iron(III)-reducing bacteria on clay minerals as the sole electron acceptor and comparison of growth yields on a variety of oxidized iron forms." <u>Applied and Environmental Microbiology</u> **68**(12): 6256-6262.
- Kostka, J. E., J. W. Stucki, et al. (2002). "Microbial reduction of Fe(III) bound in clay minerals: Laboratory investigations of growth and mineral transformation."

 <u>Abstracts of Papers of the American Chemical Society</u> **223**: U598-U598.
- Kostka, J. E., J. Wu, et al. (1999). "The impact of structural Fe(III) reduction by bacteria on the surface chemistry of smectite clay minerals." <u>Geochimica Et</u> Cosmochimica Acta **63**(22): 3705-3713.
- Kovett, V. A. and J. Cullen (1965). "Some factors affecting the cyclopropane acid

- formation in Escherichia coli." <u>Biochemistry Journal</u> **96**: 771-776.
- Kowalsky, M. B., J. Chen, et al. (2006). "Joint inversion of geophysical and hydrological data for improved subsurface characterization." <u>The Leading Edge, Soc. Explor. Geophysicists</u> **25**: 730.
- Kowalsky, M. B., S. Finsterle, et al. (2005). "Estimation of field-scale soil hydraulic and dielectric parameters through joint inversion of GPR and hydrological data." <u>Water Resources Research</u> **41**(W11425): doi: 10.1029/2005WRR004237.
- Kukkadapu, R. K., J. M. Zachara, et al. (2006). "Reductive Biotransformation of Fe in Shale-Limestone Saprolite Containing Fe(III) Oxides and Fe(II)/Fe(III) Phyllosilicates." Geochim. Cosmochim. Acta **70**: 3662-3676.
- LaBolle, E. M. and G. E. Fogg (2001). "Role of molecular diffusion in contaminant migration and recovery in an alluvial aquifer system." <u>Transport in Porous Media</u> **42**(1-2): 155-179.
- Lack, J. G., S. K. Chaudhuri, et al. (2002). "Immobilization of radionuclides and heavy metals through anaerobic bio-oxidation of Fe(II)." <u>Applied and Environmental Microbiology</u> **68**(6): 2704-2710.
- Larsen, O. and C. B. Koch (2000). "Application of 57Fe-enriched synthetic ferrihydrite to speciate the product of bacterial reduction." <u>Hyperfine Interactions</u> **126**: 225-234.
- Liger, E., L. Charlet, et al. (1999). "Surface catalysis of uranium(VI) reduction by iron(II)." <u>Geochimica Et Cosmochimica Acta</u> **63**(19-20): 2939-2955.
- Linde, H., S. Finsterle, et al. (2006). "Inversion of hydrological tracer test data using tomographic constraints." <u>Water Resources Research</u> **in press**.
- Lipton, M. S., L. Pasa-Tolic, et al. (2002). "Global analysis of the Deinococcus radiodurans proteome by using accurate mass tags." <u>Proceedings of the National</u> Academy of Sciences of the United States of America **99**(17): 11049-11054.
- Liu, C. X., Y. A. Gorby, et al. (2002). "Reduction kinetics of Fe(III), Co(III), U(VI) Cr(VI) and Tc(VII) in cultures of dissimilatory metal-reducing bacteria." <u>Biotechnology</u> and <u>Bioengineering</u> **80**(6): 637-649.
- Liu, C. X., S. Kota, et al. (2001). "Kinetic analysis of the bacterial reduction of goethite." <u>Environmental Science & Technology</u> **35**(12): 2482-2490.
- Liu, C. X., J. M. Zachara, et al. (2005). "Influence of sediment bioreduction and reoxidation on uranium sorption." <u>Environmental Science & Technology</u> **39**(11): 4125-4133.
- Long, P., Y.-j. Chang, et al. (2005). Microbial Incorporation of 13C Labeled Acetate at the Field Scale: Detection of Microbes Responsible for Reduction of U(VI). <u>The 105th General Meeting of the American Society for Microbiology</u>. Atlanta, GA.
- Lovley, D. R., F. H. Chapelle, et al. (1994). "Use of Dissolved H(2) Concentrations to Determine Distribution of Microbially Catalyzed Redox Reactions in Anoxic Groundwater." Environmental Science & Technology **28**(7): 1205-1210.
- Lovley, D. R. and S. Goodwin (1988). "Hydrogen concentrations as an indicator of the predominant terminal electron-accepting reactions in aquatic sediments."

 Geochim. Cosmochim. Acta **52**: 2993-3003.
- Lovley, D. R. and E. J. P. Phillips (1987). "Rapid Assay for Microbially Reducible Ferric Iron in Aquatic Sediments." <u>Applied and Environmental Microbiology</u> **53**(7): 1536-1540.
- Lovley, D. R. and E. J. P. Phillips (1988). "Novel Mode of Microbial Energy-Metabolism -

- Organic-Carbon Oxidation Coupled to Dissimilatory Reduction of Iron or Manganese." <u>Applied and Environmental Microbiology</u> **54**(6): 1472-1480.
- Lovley, D. R. and E. J. P. Phillips (1992). "Bioremediation of Uranium Contamination with Enzymatic Uranium Reduction." <u>Environmental Science & Technology</u> **26**(11): 2228-2234.
- Lovley, D. R. and E. J. P. Phillips (1992). "Reduction of Uranium by Desulfovibrio-Desulfuricans." <u>Applied and Environmental Microbiology</u> **58**(3): 850-856.
- Lovley, D. R., E. J. P. Phillips, et al. (1991). "Microbial Reduction of Uranium." <u>Nature</u> **350**(6317): 413-416.
- Lovley, D. R., E. E. Roden, et al. (1993). "Enzymatic Iron and Uranium Reduction by Sulfate-Reducing Bacteria." <u>Marine Geology</u> **113**(1-2): 41-53.
- Maidak, B. L., G. J. Olsen, et al. (1997). "The RDP (Ribosomal Database Project)." Nucleic Acids Research **25**(1): 109-110.
- Masselon, C., L. Pasa-Tolic, et al. (2005). "Targeted comparative proteomics by liquid chromatography-tandem Fourier ion cyclotron resonance mass spectrometry." Analytical Chemistry **77**(2): 400-406.
- Methe, B. A., K. E. Nelson, et al. (2003). "Genome of Geobacter sulfurreducens: metal reduction in subsurface environments." Science **302**: 1967-1969.
- Morrison, S. J., R. R. Spangler, et al. (1996). "Subsurface injection of dissolved ferric chloride to form a chemical barrier: Laboratory investigations." <u>Ground Water</u> **34**(1): 75-83.
- Morrison, S. J., R. R. Spangler, et al. (1995). "Adsorption of Uranium(Vi) on Amorphous Ferric Oxyhydroxide at High-Concentrations of Dissolved Carbon(Iv) and Sulfur(Vi)." Journal of Contaminant Hydrology **17**(4): 333-346.
- N'Guessan, H. A. Vrionis, et al. (2006). Surprising long-term removal of uranium from contaminated groundwater in the absence of acetate addition. <u>The 106th General Meeting of the American Society for Microbiology</u>. Orlando, FL.
- Norbeck, A. D., M. E. Monroe, et al. (2005). "The utility of accurate mass and LC elution time information in the analysis of complex proteomes." <u>Journal of the American</u> Society for Mass Spectrometry **16**(8): 1239-1249.
- Ntarlagiannis, D., K. H. Williams, et al. (2006). "Low frequency electrical response to microbial induced sulfide precipitation." <u>Journal of Geophysical Research</u> **110**(G02009): doi:10.1029/2005JG000024.
- Olsen, G. J., H. Matsuda, et al. (1994). "Fastdnaml a Tool for Construction of Phylogenetic Trees of DNA-Sequences Using Maximum-Likelihood." <u>Computer Applications in the Biosciences</u> **10**(1): 41-48.
- Ortiz-Bernad, I., R. T. Anderson, et al. (2004). "Resistance of solid-phase U(VI) to microbial reduction during in situ bioremediation of uranium-contaminated groundwater." <u>Applied and Environmental Microbiology</u> **70(12)**: 7558-7560.
- Park, S. S. and P. R. Jaffe (1996). "Development of a sediment redox potential model for the assessment of postdepositional metal mobility." <u>Ecological Modelling</u> **91**(1-3): 169-181.
- Parker, J. H., G. A. Smith, et al. (1982). "Sensitive Assay, Based on Hydroxy Fatty-Acids from Lipopolysaccharide Lipid-a, for Gram-Negative Bacteria in Sediments." <u>Applied and Environmental Microbiology</u> **44**(5): 1170-1177.
- Payne, T. E., J. A. Davis, et al. (1996). "Uranium adsorption on ferrihydrite Effects of

- phosphate and humic acid." Radiochimica Acta 74: 239-243.
- Peacock, A. D., Y. J. Chang, et al. (2004). "Utilization of microbial biofilms as monitors of bioremediation." <u>Microbial Ecology</u> **47**(3): 284-292.
- Qafoku, N. P., J. M. Zachara, et al. (2005). "Kinetic desorption and sorption of U(VI) during reactive transport in a contaminated Hanford sediment." <u>Environmental-Science & Technology</u> 39(9): 3157-3165.
- Rabouille, C. and J. F. Gaillard (1991). "A Coupled Model Representing the Deep-Sea Organic-Carbon Mineralization and Oxygen-Consumption in Surficial Sediments." Journal of Geophysical Research-Oceans **96**(C2): 2761-2776.
- Ram, R. J., N. C. VerBerkmoes, et al. (2005). "Community proteomics of a natural microbial biofilm." <u>Science</u> **308**(5730): 1915-1920.
- Rao, R. L. N., S. Sethuram, et al. (2000). "Geophysical signatures of a fracture controlled U-mineralisation: A case study from mulapalle area, Cuddapah District, Andhra Pradesh." <u>Journal of the Geological Society of India</u> **55**(4): 421-429.
- Reguera, G., K. D. McCarthy, et al. (2005). "Extracellular electron transfer via microbial nanowires." Nature **435**(7045): 1098-1101.
- Rittmann, B. E. and P. L. McCarty (2001). <u>Environmental biotechnology: principles and applications</u>. Boston, McGraw-Hill.
- Roden, E. E. and T. D. Scheibe (2005). "Conceptual and numerical model of uranium(VI) reductive immobilization in fractured subsurface sediments." Chemosphere **59**(5): 617-628.
- Roden, E. E. and T. D. Scheibe (2005). "Conceptual and numerical model of uranium(VI) reductive immobilization in fractured subsurface sediments." Chemosphere **59**(5): 617-628.
- Saripalli, K. P., C. F. Brown, et al. (2005). "Development of cellular absorptive tracers for quantitative characterization of microbial mass in flow systems." Research in Microbiology **156**(7): 790-792.
- Scheibe, T. D., Y. J. Chien, et al. (2001). "Use of quantative models to design microbial transport experiments in a sandy aquifer." <u>Ground Water</u> **39**(2): 210-222.
- Scheibe, T. D., Y. Fang, et al. (2006). "Transport and biogeochemical reactions of metals in a chemically heterogeneous aquifer." <u>Geosphere</u> in press.
- Schiewer, S. and B. Volesky (2000). Biosorption processes for heavy metal removal. <u>Environmental Microbe-Metal Interactions</u>. D. R. Lovley. Washington DC, ASM Press: 329-362.
- Scott, R. (1995). "In-Situ Bioremediation When Does It Work." <u>Journal of Environmental Management</u> **43**(2): 193-193.
- Semprini, L. and P. L. Mccarty (1992). "Comparison between Model Simulations and Field Results for Insitu Biorestoration of Chlorinated Aliphatics .2. Cometabolic Transformations." <u>Ground Water</u> **30**(1): 37-44.
- Shelobolina, E. S., R. T. Anderson, et al. (2004). "Importance of clay size minerals for Fe(III) respiration in a petroleum-contaminated aquifer." Geobiology **2**: 67-76.
- Smith, R. D., G. A. Anderson, et al. (2002). "An accurate mass tag strategy for quantitative and high-throughput proteome measurements." <u>Proteomics</u> **2**(5): 513-523.
- Smith, S. L. and P. R. Jaffe (1998). "Modeling the transport and reaction of trace metals

- in water-saturated soils and sediments." <u>Water Resources Research</u> **34**(11): 3135-3147.
- Suzuki, Y., S. D. Kelly, et al. (2002). "Radionuclide contamination Nanometre-size products of uranium bioreduction." Nature **419**(6903): 134-134.
- Suzuki, Y., S. D. Kelly, et al. (2005). "Direct microbial reduction and subsequent preservation of uranium in natural near-surface sediment." <u>Applied and Environmental Microbiology</u> **71**(4): 1790-1797.
- Tokunaga, T. K., J. M. Wan, et al. (2005). "Uranium reduction in sediments under diffusion-limited transport of organic carbon." <u>Environmental Science & Technology</u> **39**(18): 7077-7083.
- Tollefson, T. S. and R. B. McKercher (1983). "The degradation of 14C-labelled phosphatidyl choline in soil." <u>Soil Biol. Biochem</u> **15**: 145-148.
- Tyson, G. W., J. Chapman, et al. (2004). "Community structure and metabolism through reconstruction of microbial genomes from the environment." <u>Nature</u> **428**(6978): 37-43.
- Urrutia, M. M., E. E. Roden, et al. (1999). "Influence of aqueous and solid-phase Fe(II) complexants on microbial reduction of crystalline iron(III) oxides." <u>Environmental Science & Technology</u> **33**(22): 4022-4028.
- VerBerkmoes, N. C., J. L. Bundy, et al. (2002). "Integrating "top-down" and "bottom-up" mass spectrometric approaches for proteomic analysis of Shewanella oneidensis." Journal of Proteome Research 1(3): 239-252.
- VerBerkmoes, N. C., M. B. Shah, et al. (2006). "Determination and comparison of the baseline proteomes of the versatile microbe Rhodopseudomonas palustris under its major metabolic states." <u>Journal of Proteome Research</u> **5**(2): 287-298.
- Vrionis, H. A., R. T. Anderson, et al. (2005). "Microbiological and geochemical heterogeneity in an in situ uranium bioremediation field site." <u>Applied and Environmental Microbiology</u> **71**(10): 6308-6318.
- Waite, T. D., J. A. Davis, et al. (1994). "Uranium(Vi) Adsorption to Ferrihydrite Application of a Surface Complexation Model." <u>Geochimica Et Cosmochimica Acta</u> **58**(24): 5465-5478.
- Wan, J. M., T. K. Tokunaga, et al. (2005). "Reoxidation of bioreduced uranium under reducing conditions." <u>Environmental Science & Technology</u> **39**(16): 6162-6169.
- Wan, X., N. C. VerBerkmoes, et al. (2004). "Transcriptomic and Proteomic Characterization of the Fur Modulon in the Metal-Reducing Bacterium Shewanella oneidensis." <u>Journal of Bacteriology</u> **186**: 8385-8400.
- Wang, S., P. R. Jaffe, et al. (2003). "Simulating bioremediation of uranium-contaminated aquifers; uncertainty assessment of model parameters." <u>Journal of Contaminant</u> Hydrology **64**(3-4): 283-307.
- White, D. (1995). "Chemical ecology: Possible linkage between macro-and microbial ecology." Oikos **74**: 174-181.
- White, D., J. Stair, et al. (1996). "Quantitative Comparisons of in situ Microbial Biodiversity by Signature Biomarker Analysis." <u>J. Indust. Microbiol.</u> **17**: 185-196.
- White, D. C. (1988). "Validation of quantitative analysis for microbial biomass, community structure, and metabolic activity." Adv. Limnol. **31**: 1-18.
- White, D. C., W. M. Davis, et al. (1979). "Determination of the Sedimentary Microbial Biomass by Extractable Lipid Phosphate." <u>Oecologia</u> **40**(1): 51-62.

- White, D. C. and P. E. Long. (2006). Lipid analysis of microbial processes and communities at a Uranium Bio-immobilization site Using 13C-labled Acetate Amendment. <u>Department of Energy-ERSD PI Meeting</u>. Warrenton, VA.
- Widdowson, M. A., F. J. Molz, et al. (1988). "A Numerical Transport Model for Oxygen-Based and Nitrate-Based Respiration Linked to Substrate and Nutrient Availability in Porous-Media." <u>Water Resources Research</u> **24**(9): 1553-1565.
- Williams, K. H., A. Kemna, et al. (2006). The Role of Geoelectrical Methods in Monitoring Stimulated Sulfate-Reduction: Insights Gained From Field-Scale Experiments. <u>AGU Fall Meet. Suppl.</u>, Eos Trans. 87(52) Abstract NS21A-06
- Williams, K. H., D. Ntarlagiannis, et al. (2005). "Geophysical imaging of stimulated microbial biomineralization." <u>Environmental Science & Technology</u> **39**(19): 7592-7600.
- Wu, Y., L. D. Slater, et al. (2006). "Low Frequency Electrical Properties of Corroded Iron Barrier Cores." <u>Environmental Science & Technology</u> **40**: 2254-2261.
- Yabusaki, S., K. Cantrell, et al. (2001). "Multicomponent reactive transport in an in situ zero-valent iron cell." <u>Environmental Science & Technology</u> **35**(7): 1493-1503.
- Yanase, N., T. E. Payne, et al. (1995). "Groundwater Geochemistry in the Koongarra Ore Deposit, Australia .1. Implications for Uranium Migration." <u>Geochemical Journal</u> **29**(1): 1-29.
- Yang, J. B. and B. Volesky (1999). "Biosorption of uranium on Sargassum biomass." <u>Water Research</u> **33**(15): 3357-3363.
- Zachara, J. M., R. K. Kukkadapu, et al. (2004). "Biogeochemical transformation of Fe minerals in a petroleum-contaminated aquifer." <u>Geochimica Et Cosmochimica Acta</u> **68**(8): 1791-1805.